

Bibliography

1. Ahn, Y.O., et al. Exogenous sucrose utilization and starch biosynthesis among sweet potato cultivars, *Carbohydr. Res.* **345** (1), 55--60, 2010.
2. Noh, S.A., et al. *SRDI* is involved in the auxin-mediated initial thickening growth of storage root by enhancing proliferation of metaxylem and cambium cells in sweetpotato (*Ipomoea batatas*), *J. Exp. Bot.* **61**, 1337--1349, 2010.
3. Mohan, C. Tropical Tuber Crops, in *Advances in Horticulture Biotechnology: Molecular Markers Assisted Selection-Vegetables, Ornamentals and Tuber Crops*, H.P. Singh et al, eds., Westville Publishing House, New Delhi (India), 2011, 187-230.
4. Anbuselvi, S., et al. A comparative study on biochemical constituents of sweet potatoes from Orissa and Tamilnadu and its curd formation, *J. Chem. Pharm. Res.* **4** (11), 4879--4882, 2012.
5. Senanayake, S.A., et al. Comparative analysis of nutritional quality of five different cultivars of sweet potatoes (*Ipomea batatas* (L) Lam) in Sri Lanka, *Food Science & Nutrition* **1**, 284--291, 2013.
6. Loebenstein, G. & Thottappilly, G. *The Sweetpotato*. Springer Sciences Business Media BV, eds. Dordrecht, Netherlands, 2009.
7. Teow, C.C., et al. Antioxidant activities, phenolic and β -carotene contents of sweet potato genotypes with varying flesh colours, *Food Chem.* **103**, 829--838, 2007.
8. Scott, G.J., et al. Global projections for root and tuber crops to the year 2020, *Food Policy* **25**, 561--597, 2000.
9. Diop, A. Storage and processing of roots and tubers in the tropics, in *Food and Agriculture Organization of the United Nations, Agro-Industries and Post-Harvest Management Service*, D.J.B. Calverley ed., Agricultural Support Systems Division. Food and Agriculture Organization, Rome, Italy, 1998, 38-50.
10. Jiang, X., Jianjun, H., & Wang, Y. (2004a). Sweetpotato processing and product research and development at the Sichuan Academy of Agricultural Sciences. In “Sweetpotato post-Harvest Research and Development in China” (K.O. Fuglie and M. Hermann, eds), Proceedings of an International Workshop held in Chengdu, Sichuan, PR China, November 7--8, 2001, International Potato Center (CIP), Bogor, Indonesia.
11. Woolfe, J.A. *Sweet potato: an untapped food resource*, Cambridge, UK: Cambridge University press, 1992.

12. Ishida, H., et al. Nutritive evaluation on chemical components of leaves, stalks and stems of sweetpotatoes (*Ipomoea batatas* poir), *Food Chem.* **68**, 359--367, 2000.
13. Antonio, G.C., et al. Sweet Potato: Production, Morphological and Physicochemical Characteristics, and Technological Process. In: Focus on sweet potato, *Fruit, Vegetable and Cereal Science and Biotechnology* **5** (Special Issue 2), 1--18, 2011.
14. Oke, M.O., & Workneh, T.S. A review on sweet potato postharvest processing and preservation technology, *Afr. J. Agric. Res.* **8** (40), 4990—5003, 2013.
15. Sosinski, B., He, J., Cervantes-Flores, R., Pokrzywa, M., Bruckner, A. & Yencho, G.C. Sweet potato genomics at North Carolina state University. Ames, T (ed). Proceedings of the first International Conference on sweet potato. Food and Health for the future, *Acta Horticult.* **583**, 69--76, 2002.
16. Wanda, C.W. Genetic improvement for meeting human nutrition needs. Quebedeaux, B. and Bliss F (editors). Proceedings of the first International symposium on horticulture and human nutrition, Contributor of fruits and vegetables. Prentice Hall. pp. 191--199, 1987.
17. Bovell-Benjamin, A.C. Sweet potato: a review of its past, present, and future role in human nutrition, *Adv. Food Nutr. Res.* **52**, 1--59, 2007.
18. Huo, G., Lin, S. & Green, S. *Sweet potato germplasm for international cooperation.* International Cooperation's Guide, AVRDC, 1985.
19. Chen, Z., et al. Evaluation of starch noodles made from three typical Chinese sweetpotato starches, *J. Food Sci.* **67** (9), 3342--3347, 2002.
20. Moorthy, S. N. Physicochemical and functional properties of tropical tuber starches: A review, *Starch/Starke* **54** (12), 559--592, 2002.
21. Hoover, R. Composition, molecular structure, and physicochemical properties of tuber and root starches: a review, *Carbohydr. Polym.* **45** (3), 253--267, 2001.
22. Valetudie, J.C., et al. Influence of cooking procedures on structure and biochemical changes in sweet potato, *Starch/Starke* **51** (11-12), 389--397, 1999.
23. Walter, W.M., et al. Sweet potato protein: a review, *J. Agric. Food Chem.* **32**, 695--699, 1984.

24. Diop, A. Storage and processing of roots and tubers in the tropics, in *Food and Agriculture Organization of the United Nations, Agro-Industries and Post-Harvest Management Service*, D.J.B. Calverley, ed., Food and Agriculture Organization, Rome, Italy 1998, 38–50.
25. Tewe, O.O., Ojeniyi, F.E. & Abu, O.A. *Sweetpotato Production, Utilization and Marketing in Nigeria*. Social Sciences Department, International Potato Center (CIP), Lima, Peru, 2003.
26. Bradbury, J.H. Chemical Composition of cooked and uncooked sweet potato and its significance for human nutrition, in *Sweet potato research and development for small farmers*, K.T. Mackay et al, eds, SEAMEO-SEARCA, Philippines, 1989, 213–225.
27. Ravindran, V., et al. Biochemical and nutritional assessment of tubers from 16 cultivars of sweet potato (*Ipomoea batatas* L.), *J. Agric. Food Chem.* **43**, 2646--2651, 1995.
28. Wanda, & Collins W. Genetic improvement for meeting human Nutrition needs. Quebedeaux, B and Bliss, F (Editors). Proceedings of the first international symposium on horticulture and human nutrition, Contributor of fruits and vegetable, Prentice Hall, 191–199, 1987.
29. Guillon, F., & Champ, M. Structural and physical properties of dietary fibres, and consequences of processing on human physiology, *Food Res. Int.* **33**, 233--245, 2000.
30. Brody, T. *Nutritional Biochemistry*, Academic Press, San Diego, CA, 1994.
31. Huang, A.S., et al. Content of alpha-, beta-carotene, and dietary fiber in 18 sweetpotato varieties grown in Hawaii, *J. Food Compost. Anal.* **12**, 147--151, 1999.
32. Oboh, S., et al. Some aspects of the biochemistry and nutritional value of the sweetpotato (*Ipomoea batatas*), *Food Chem.* **31**, 9--18, 1989.
33. Walter Jr., W. M., & Catignani, G. L. Biological Quality and Composition of Sweet Potato Protein Fractions, *J. Agric. Food Chem.* **29** (4), 797--799, 1981.
34. Rose, I.M., & Vasanthakalam, H. Comparison of the Nutrient composition of four sweet potato varieties cultivated in Rwanda. *Am. J. Food Nutr.* **1**, 34--38, 2011.
35. Olaofe, O., & Sanni, C.O. Mineral contents of agricultural products, *Food Chem.* **30**, 73--77, 1988.

36. Makki, H.M., et al. Chemical composition of Egyptian sweetpotato, *Food Chem.* **20**, 39--44, 1986.
37. Islam, S., et al. Identification and characterization of foliar polyphenolic composition in sweetpotato (*Ipomoea batatas* L.) genotypes, *J. Agric. Food Chem.* **50**, 3718--3722, 2002.
38. Plata, N., et al. Effect of methyl jasmonate and p-coumaric acid on anthocyanin composition in a sweetpotato cell suspension culture, *Biochem. Eng. J.* **14**, 171--177, 2003.
39. Knaes. (Kyushu National Agricultural Experiment Station), No. 1, December 1995, Tokyo, Japan, 1995.
40. Cevallos-Casals, B.A., & Cisneros-Zevallos, L.A. Bioactive and functional properties of purple sweetpotato (*Ipomoea batatas* [L.] Lam), *Acta Hort. (ISHS)* **583**, 195--203, 2002.
41. Otake, K., et al. Chemical structures of two anthocyanins from purple sweet potatoes *Ipomoea batatas*, *Phytochemistry* **31** (6), 2127--2130, 1992.
42. Terahara, N., et al. Six diacylated anthocyanins from storage roots of purple sweet potato, *Ipomoea batatas*, *Biosci. Biotech. Biochem.* **63** (8), 1420--1424, 1999.
43. Terahara, N., et al. Anthocyanins in callus induced from purple storage root of *Ipomoea batatas* L. *Phytochemistry* **54**, 919--922, 2000.
44. Terahara, N., et al. Characterization of acylated anthocyanins in callus induced from storage root of purple-fleshed sweet potato, *Ipomoea batatas* L. *J. Biomed. Biotechnol.* **5**, 279--286, 2004.
45. Ameny, M.A., & Wilson, P.W. Relationship between Hunter Color Values and β -Carotene Contents in White-Fleshed African Sweetpotatoes (*Ipomoea batatas* Lam), *J. Sci. Food Agr.* **73**, 301--306, 1997.
46. Koala, M., et al. Evaluation of eight orange fleshed sweet potato (OFSP) varieties for their total antioxidant, total carotenoid and polyphenolic contents, *Journal of Natural Sciences Research* **3**, 67--72, 2013.
47. Manrique, K., & Hermann, M. Effects of GxE Interaction on Root yield and β -carotene content of selected sweet potato (*Ipomoea batatas* (L.) Lam) varieties and breeding clones, *CIP Program Report 1999--2000*, 281--287, 2001.
48. Cinar, I. Effects of cellulose and pectinase concentrations on the colour yield of enzyme extracted plant carotenoids, *Process Biochem.* **40**, 945--949, 2005.

49. USDA (U.S. Department of Agriculture), Agricultural Research Service. (2009). USDA National Nutrient Database for Standard Reference, Release 22. Nutrient Data Laboratory Home Page, <http://www.ars.usda.gov/ba/bhnrc/ndl>, Accessed 14 September 2012.
50. Konczak-Islam, I., et al. Potential chemopreventive properties of anthocyanin-rich aqueous extracts from *in vitro* produced tissue of sweetpotato, *J. Agric. Food Chem.* **51**, 5916--5922, 2003.
51. Suda, I., et al. Physiological functionality of purple-fleshed sweetpotatoes containing anthocyanins and their utilization in foods, *JPN. AGR. RES. Q.* **37**, 167--173, 2003.
52. Dreher, D., & Junod, A.F. Role of oxygen free radicals in cancer development, *Eur. J. Cancer* **32A**, 30--38, 1996.
53. Scheibmeir, H.D., et al. A review of free radicals and antioxidants for critical care nurses, *Intensive Crit. Care Nurs.* **21**, 24--28, 2005.
54. Yoshimoto, M., et al. Antimutagenicity of sweetpotato (*Ipomoea batatas*) roots, *Biosci. Biotechnol. Biochem.* **63** (3), 536--541, 1999.
55. Yoshimoto, M., et al. Antimutagenicity of deacylated anthocyanins in purple-fleshed sweetpotato, *Biosci. Biotechnol. Biochem.* **65** (7), 1652--1655, 2001.
56. Matsui, T., et al. Anti-hyperglycemic effect of diacylated anthocyanins derived from *Ipomoea batatas* cultivar ayamurasaki can be achieved through the α -glucosidase inhibitory action, *J. Agric. Food Chem.* **50**, 7244--7248, 2002.
57. Islam, S. Sweetpotato (*Ipomoea batatas* L.) leaf: its potential effect on human health and nutrition, *J. Food Sci.* **71** (2), 13--21, 2006.
58. Weiss, R.F. & Finkelmann, A. Herbal Medicine, 2nd ed. Thieme, Stuttgart, 2000.
59. Zhang, K., et al. ISSR-Based Molecular Characterization of an Elite Germplasm Collection of Sweet Potato (*Ipomoea batatas* L.) in China, *Journal of Integrative Agriculture Advanced* 1--18, 2014.
60. FAO (Food and Agricultural Organization). Production and harvesting areas of potato and sweet potato crops. Stat Data 2005, 2013 FAO, Rome, Italy: Available from www.fao.org.

61. Rukundo, P., et al. Storage root formation, dry matter synthesis, accumulation and genetics in sweetpotato, *Aust. J. Crop Sci.* **7**, 2054--2061, 2013.
62. Lemoine, R., et al. Source-to-sink transport of sugar and regulation by environmental factors, *Front. Plant Sci.* **4**, 272, 2013.
63. Lowe, S.B., & Wilson, L.A. Comparative analysis of tuber development in six sweet potato (*Ipomoea batatas* (L.) Lam.) Cultivars. 1. Tuber initiation, tuber growth and partition of assimilate, *Ann. Bot.* **38**, 307--17, 1974.
64. Kays, S.J. The physiology of yield in the sweetpotato, in. *Sweet potato products: a natural resource for the tropics*, J.C. Bouwkamp eds., CRC Press. Boca Raton, FL, 1985, 79-132.
65. Ravi, V., & Indira, P. Crop physiology of sweetpotato, *Hortic. Rev.* **23**, 277--339, 1999.
66. Nakatani, M., & Komeichi, M. Changes in the endogenous level of zeatin riboside, abscisic acid and indole acetic acid during formation and thickening of tuberous roots in sweet potato, *JPN J CROP SCI* **60**, 91--100, 1991.
67. Togari, Y. A study of tuberous root formation in sweet potato, *Bull. Natl. Agric. Exp. Stn. Tokyo* **68**, 1--96, 1950.
68. Eguchi, T., et al. Growth of sweetpotato tuber as affected by the ambient humidity, *Biotronics* **27**, 93--96, 1998.
69. Hill, J., et al. Biomass accumulation in hydroponically grown sweetpotato in a controlled environment: a preliminary study, *Acta Hort.* **440**, 25--30, 1996.
70. Loretan, P.A., et al. Effects of several environmental factors on sweetpotato growth, *Adv. Space Res.* **14**, 277--280, 1994.
71. van Heerden, P.D., & Laurie, R. Effects of prolonged restriction in water supply on photosynthesis, shoot development and storage root yield in sweet potato, *Physiol. Plant* **134**, 99--109, 2008.
72. Matsuo, T., et al. Identification of free cytokinins and the changes in endogenous levels during tuber development of sweet potato (*Ipomoea batatas* Lam.), *Plant Cell Physiol.* **24**, 1305--1312, 1983.
73. Nakatani, M., & Komeichi, M. Changes in endogenous indole acetic acid level during development of roots in sweet potato, *JPN J. CROP SCI.* **61**, 683--684, 1992.

74. Eguchi, T., & Yoshida, S. Effects of application of sucrose and cytokinin to roots on the formation of tuberous roots in sweetpotato (*Ipomoea batatas* (L.) Lam.), *Plant Root* **2**, 7--13, 2008.
75. Wang, Q., et al. Endogenous hormone concentration in developing tuberous roots of different sweet potato genotypes, *AGRIC. SCI. CHINA* **5**, 919--927, 2006.
76. McDavid, C.R., & Alamu, S. The effect of growth regulators on tuber initiation and growth in rooted leaves of two sweet potato cultivars, *Ann. Bot.* **45**, 363--364, 1980.
77. Ekanayake, I.J. Evaluation of potato and sweetpotato genotypes for drought resistance. *CIP Research Guide* **19**, 1--16, 1990.
78. Anioke, S.C. Effect of time of planting and harvesting of sweetpotato (*Ipomoea batatas* L. Lam.) on yield and insect damage in south eastern Nigeria, *Entomology* **21**, 137--141, 1996.
79. Nedunchezhiyan, M., et al. Sweet Potato Agronomy, *Fruit, Veg. Cereal Sci. Biotech.* **6** (Special Issue 1), 1--10, 2012.
80. Purseglove, J.W. *Convulvulaceae In Tropical Crops. Dicotyledons*. Longmans, Green and Co. Ltd., London: 78—88, 1968.
81. Dasgupta, M., et al. Screening of sweet potato genotypes for salinity stress. in *14th Triennial Symposium of International Society of Tropical Root Crops*, Central Tuber Crops Research Institute, Thiruvananthapuram, India, 166--167, 2006.
82. Mukherjee, A., et al. Response of orange flesh sweet potato genotypes to salinity stress, in *14th Triennial Symposium of International Society of Tropical Root Crops*, 20-26 November 2006, Central Tuber Crops Research Institute, Thiruvananthapuram, India, pp 151—152, 2006.
83. Nayar, G.G. & Naskar, S.K. Varietal improvement in sweet potato, in *Advances in Horticulture: Tuber Crops* (Vol 8), K.L. Chadha et al, eds., Malhotra Publishing House, New Delhi, India, 1994, 101-112.
84. Rossell, G., et al. The sweetpotato germplasm collection at CIP, Lima-Peru. Paper presented in the *CIP-UPWARD Workshop "Sweetpotato Global Conservation Strategy"*. The Global Crop Diversity Trust. Manila, Philippines, 2007.
85. Nakashima, K., & Yamaguchi-Shinozaki, K. ABA signaling in stress-response and seed development, *Plant Cell Rep.* **32**, 959--970, 2013.

86. Kramer, P.J. & Boyer, J.S. *Water relations of plants and soils*, Academic Press, UK, 1995.
87. Anselmo, B.A., et al. Screening sweetpotato for drought tolerance in the Philippine highlands and genetic diversity among selected genotypes, *Trop. Agric.* **75**, 1998.
88. Anjum, S.A., et al. Review: Morphological, physiological and biochemical responses of plants to drought stress, *Afr. J. Agric. Res.* **6** (9), 2026--2032, 2011.
89. Jiang, Y., et al. A proteomic analysis of storage stress responses in *Ipomoea batatas* (L.) Lam. tuberous root, *Mol. Biol. Rep.* **39**, 8015--8025, 2012.
90. Shonga, E.M., et al. Review of entomological research on sweetpotato in Ethiopia. *DJFAS* **1**, 83--92, 2013.
91. Ehisianya, C.N., et al. Field efficacy of Neem seed oil and diazinon in the management of sweetpotato weevil, *Cylas puncticollis* (Boh.) in south eastern Nigeria, *J. Plant Res.* **2**, 135-144, 2013.
92. McGregor, C.E., et al. Differential gene expression of resistant and susceptible sweetpotato plants after infection with the causal agents of sweetpotato virus disease, *J. Am. Soc. Hortic. Sci.* **134**, 658--666, 2009.
93. Grüneberg, W., Mwanga, R., Andrade, M. & Espinoza, J. Selection methods: breeding clonally propagated crops, in *Plant breeding and farmer participation*, S. Ceccarelli et al, eds., Food and Agriculture Organisation of the United Nations, Rome, Italy, 2009, 275-322.
94. Martin, F.M., & Jones, A. Breeding sweetpotatoes. *Plant Breeding Review* **4**, 313--345, 1986.
95. Ames, T., Smit, N.E.J.M., Braun, A.R., O'Sullivan, J.N. & Skoglund, L.G. *Sweetpotato: Major pests, diseases, and nutritional disorders*, International Potato Center (CIP), Lima, Peru, 1996.
96. Ndunguru, J., & Kapinga, R. Viruses and virus-like diseases affecting sweetpotato subsistence farming in southern Tanzania, *Afr. J. Agric. Res.* **2**, 232--239, 2007.
97. Stover, P.J. Nutritional genomics, *Physiol. Genomics* **16**, 161--165, 2004.
98. Mutch, D.M., et al. Nutrigenomics and nutrigenetics: The emerging faces of nutrition, *FASEB J.* **19**, 1602--1616, 2005.

99. Trujillo, E., et al. Nutrigenomics, proteomics, metabolomics, and the practice of dietetics, *J. Am. Diet Assoc.* **106** (3), 403--413, 2006.
100. Sales, N.M.R., et al. Nutrigenomics: Definitions and Advances of This New Science, *Journal of Nutrition and Metabolism* **2014** (2014), 1--6, 2014.
101. Tian, L., & Della Penna, D. The promise of agricultural biotechnology for human health, *BRIT. FOOD J.* **103** (11), 777--779, 2001.
102. Zhang, X., et al. Novel omics technologies in nutrition research, *Biotechnol Adv* **26**, 169 -- 176, 2008.
103. Ganesh, V., & Hettiarachchy, N.S. Nutriproteomics: A promising tool to link diet and diseases in nutritional research, *Biochim. Biophys. Acta* **1824**, 1107--1117, 2012.
104. Mandal, S., & Mandal, R. K. Seed storage proteins and approaches for improvement of their nutritional quality by genetic engineering, *Curr. Sci.* **79**, 576--589, 2000.
105. Ufaz, S., & Galili, G. Improving the content of essential amino acids in crop plants: goals and opportunities, *Plant Physiol.* **147**, 954--961, 2008.
106. Folk, W.R. How to improve plant protein quality: err on the side of goodness. *ISB News Report*, August 2003 (<http://www.isb.vt.edu>), 2003.
107. Galili, G. Regulation of lysine and threonine synthesis, *Plant Cell* **7**, 899--906, 1995.
108. Karchi, H., et al. Seed specific expression of a bacterial desensitized aspartate kinase increases the production of seed threonine and methionine in transgenic tobacco, *Plant J.* **3**, 721--727, 1993.
109. Falco, S.C., et al. Transgenic canola and soyabean seeds with increased lysine, *Biotechnology* **13**, 577--582, 1995.
110. Randall, J., et al. A modified 10 KD zein protein produces two morphologically distinct protein bodies in transgenic tobacco, *Plant Sci.* **150**, 21--28, 2000.
111. Hoffman, L.M., et al. A modified storage protein in synthesized, processed and degraded in the seed of transgenic plants, *Plant.Mol.biol.* **11**, 717--729, 1988.
112. Chakraborty, S., et al. Increased nutritive value of transgenic potato by expressing a nonallergic seed albumin gene from *Amaranthus hypochondriacus*, *Proc. Natl. Acad. Sci.* **97**, 3724--3729, 2000.

113. Chakraborty, S., et al. Next-generation protein-rich potato expressing the seed protein gene *AmA1* is a result of proteome rebalancing in transgenic tuber, *Proc. Natl. Acad. Sci.* **107**, 17533--17538, 2010.
114. Altenbach, S.B., et al. Accumulation of a Brazil nut albumin in seeds of transgenic canola results in enhanced levels of seed protein methionine, *Plant Mol. Biol.* **18**, 235--245, 1992.
115. Sharma, S.B., et al. Expression of a sulfur-rich maize seed storage protein, δ -zein, in white clover (*Trifolium repens*) to improve forage quality, *Mol. Breed.* **4**, 435--448, 1998.
116. Melo, V.M.M., et al. Allergenicity and tolerance to proteins from Brazil nut (*Bertholletia excelsa* HBK), *Food Agricult. Immunol.* **6**, 185--195, 1994.
117. Yang, M.S., et al. Expression of a synthetic gene for improved protein quality in transformed potato plants, *Plant Sci.* **64**, 99--111, 1989.
118. Egnin, M., & Prakash, C.S. Transgenic sweetpotato expressing a synthetic storage protein gene exhibits high level of total protein and essential amino acids, *In Vitro Cell Dev. Biol.* **33**, 52A, 1997.
119. Kim, J.H., Cetiner, S. & Jaynes, J.M. Enhancing the nutritional quality of crop plants: design, construction and expression of an artificial plant storage protein gene, in *Molecular Approaches to Improving Food Quality and Safety*. D. Bhatnagar et al, eds., An avi book, New York, 1992, 1—36
120. Agros, P.K., et al. A structural model for maize zein proteins, *J. Biol. Chem.* **257**, 9984--9990, 1982.
121. Agrawal, L., et al. Comparative proteomics reveals a role for seed storage protein, AmA1 in cellular growth, development and nutrient accumulation, *J. Proteome. Res.* **12**, 4904--4930, 2013.
122. Millerd, A. Biochemistry of Legume Seed Proteins, *Annu. Rev. Plant Physiol.* **26**, 53--72, 1975.
123. Osborne, T. B. *The Vegetable Proteins*, Longmans Green, London, 1924.
124. Larkins, B. A. Genetic engineering of plants: An agricultural perspective, Plenum, New York), 1983.
125. Hoffman, L. M., et al. A modified storage protein is synthesized, processed, and degraded in the seeds of transgenic plants, *Plant Mol. Biol.* **11**, 717--729, 1988.

126. Guerche, P., et al. Expression of the 2S albumin from *Bertholletia excelsa* in *Brassica napus*, *Mol. Gen. Genet.* **221**, 306--314, 1990.
127. Raina, A., & Datta, A. Molecular cloning of a gene encoding a seed-specific protein with nutritionally balanced amino acid composition from *Amaranthus*, *Proc. Natl. Acad. Sci.* **89**, 11774--11778, 1992.
128. Ling, L.J., et al. Expression and characterization of two domains of *Pinellia ternata* agglutinin (PTA), a plant agglutinin from *Pinellia ternata* with antifungal activity, *World J. Microb Biot.* **26**, 545--554, 2010.
129. Jin, S., et al., *Pinellia ternata* agglutinin expression in chloroplasts confers broad spectrum resistance against aphid, whitefly, *Lepidopteran* insects, bacterial and viral pathogens, *Plant Biotechnol. J.* **10**, 313--327, 2012.
130. Kuiper, H.A. Assessment of the food safety issues related to genetically modified foods, *Plant J.* **27** (6), 503--528, 2001.
131. Kiambi, D.K., et al. Linking transcript profiles to metabolites and metabolic pathways: A systems biology approach to transgene risk assessment, *Plant Omics Journal* **1** (1), 26--36, 2008.
132. Iglesias, V.A., et al. Molecular and cytogenetic analyses of stably and unstably expressed transgene loci in tobacco, *Plant Cell* **9**, 1251--1264, 1997.
133. Pedersen, C., et al. Localization of introduced genes on the chromosomes of transgenic barley, wheat, triticale by fluorescence *in situ* hybridization, *Theor. Appl. Genet.* **94**, 749--757, 1997.
134. Spertini, D., et al. Screening of transgenic plants by amplification of unknown genomic DNA flanking T-DNA, *Biotechniques* **27**, 308--314, 1999.
135. Thomas, W.T.B., et al. Identification of a QTL decreasing yield in barley linked to Mlo powdery mildew resistance, *Mol. Breed.* **4**, 381--393, 1998.

136. Schena, M., et al. Quantitative monitoring of gene expression patterns with a complementary DNA microarray, *Science* **270**, 467--470, 1995.
137. Schena, M., et al. Parallel human genome analysis: microarray-based expression monitoring of 1000 genes, *Proc. Natl. Acad. Sci.* **93**, 10614--10619, 1996.
138. Kuiper, H.A., et al. Exploitation of molecular profiling techniques for GM food safety assessment, *Curr. Opin. Biotechnol.* **14**, 238--243, 2003.
139. Baudo, M. M., et al. Transgenesis has less impact on the transcriptome of wheat grain than conventional breeding, *Plant Biotechnol. J.* **4**, 369--380, 2006.
140. Deepak, S.A., et al. Real-time PCR: revolutionizing detection and expression analysis of genes, *Curr. Genomics* **8**, 234--251, 2007.
141. Schena, M., et al. Quantitative monitoring of gene expression patterns with a complementary DNA microarray, *Science* **270**, 467--470, 1995.
142. Anderson, L., & Seilhamer J. A comparison of selected mRNA and protein abundances in human liver, *Electrophoresis* **18**, 533--537, 1997.
143. Gygi, S.P., et al. Correlation between Protein and mRNA Abundance in Yeast, *Mol. Cell. Biol.* **19**, 1720--1730, 1999.
144. Greenbaum, D., et al. Comparing protein abundance and mRNA expression levels on a genomic scale, *Genome Biol.* **4**, 117--125, 2003.
145. Pisitkun, T., et al. Tandem mass spectrometry in physiology, *Physiology (Bethesda)* **2**, 390--400, 2007.
146. O'Farrell, P. H. High resolution two-dimensional electrophoresis of proteins, *J. Biol. Chem.* **250**, 4007--4021, 1975.
147. Berkelman, T., & Stenstedt, T. 2-D Electrophoresis using immobilized pH gradient: Principles and methods, *Amersham Biosciences*, 80-6429-60, 1998.
148. Subba, P., et al. Characterization of the nuclear proteome of a dehydration-sensitive cultivar of chickpea and comparative proteomic analysis with a tolerant cultivar, *Proteomics* **13**, 1973--1992, 2013.

149. Jaiswal, D. et al. Comparative proteomics of dehydration response in the rice nucleus: new insights into the molecular basis of genotype specific adaptation, *Proteomics* **13**, 3478-3497, 2013.
150. MacBeath, G., & Schreiber, S.L. Printing proteins as microarrays for high-throughput function determination, *Science* **289**, 1760--1763, 2000.
151. Pandey, A., & Mann, M. Proteomics to study genes and genomes, *Nature* **405**, 837--846, 2000.
152. Machuka, J., & Okeola, O.G. One- and two-dimensional gel electrophoretic identification of African yam bean seed proteins, *J. Agric. Food Chem.* **48**, 2296--2299, 2000.
153. Fraser, P.D., et al. Application of high-performance liquid chromatography with photodiode array detection to the metabolic profiling of plant isoprenoids, *Plant J.* **24**, 551-558, 2000.
154. Noteborn, H.P.J.M., et al. Chemical fingerprinting for the evaluation of unintended secondary metabolic changes in transgenic food crops, *J. Biotechnol.* **77**, 103--114, 2000.
155. Sihachakr, D. & Ducreux, G. Regeneration of plants from protoplasts of sweet potato (*Ipomoea batatas* L. Lam.), in *Plant Protoplasts and Genetic Engineering IV*, in: *Biotechnology in Agriculture and Forestry*, 23, Y.P.S. Bajaj eds., Springer-Verlag, Berlin, Heidelberg, 1993, 43-59.
156. González, R.G., et al. Efficient regeneration and *Agrobacterium tumefaciens* mediated transformation of recalcitrant sweet potato (*Ipomoea batatas* L.) cultivars, *Asia Pac. J. Mol. Biol. Biotechnol.* **16**, 25--33, 2008.
157. Dhir, S.K, et al. Plant regeneration via somatic embryogenesis, and transient gene expression in sweet potato protoplasts, *Plant Cell Rep.* **17**, 665--669, 1998.
158. Jansson, R. K. & Raman, K. V. Sweet potato pest management: a global overview, in *Sweet potato pest management: a global perspective*. R.K. Jansson et al, eds., Westview, Boulder, 1991, 1-12.

159. Lowe, J.M., Hamilton, W.D.O. & Newell, C.A. Genetic transformation in *Ipomoea batatas* (L.) Lam. (Sweet potato), in *Biotechnology in Agriculture and Forestry. 29: Plant Protoplasts and Genetic Engineering V*, Y.P.S. Bajaj eds. Springer, Heidelberg, 1994, 304-316.
160. Dhir, S. K., Singh H.P. & Dhir, S. Sweet Potato, in *Compendium of Transgenic Crop Plants: Transgenic Sugar, Tuber and Fiber Crops*, C. Kole et al, eds., Blackwell Publishing Ltd, Glasgow, UK, 2009, 157-176.
161. Al-Juboory, K.H., & Skirvin, R.M. *In vitro* regeneration of *Agrobacterium*-transformed sweet potato (*Ipomoea batatas* L.), *Plant Growth Regulator Science of American Quarterly* **19**, 82--89, 1991.
162. Chee, R.P., & Cantliffe, D.J. Improved production procedures for somatic embryos of sweet potato for a synthetic seed system, *HortScience* **27**, 1314--1316, 1992.
163. Prakash, C.S. Sweet potato biotechnology: progress and potential, *Biotechnol. Dev. Monit*, **18**, 1819--1822, 1994.
164. Jarret, R.L., et al. Somatic embryogenesis in sweet potato, *HortScience* **19**, 397--398, 1984.
165. Chee, R.P., et al. Optimizing embryogenic callus and embryo growth of a synthetic seed system for sweet potato by varying media nutrient concentrations, *J. Am. Soc. Hort. Sci.* **117**, 663--667, 1992.
166. Desamero, N.V., et al. Picolinic acid induced direct somatic embryogenesis in sweet potato, *Plant Cell Tiss. Organ. Cult.* **37**, 103--110, 1994.
167. Gosukonda, R.M., et al. Shoot regeneration *in vitro* from diverse genotypes of sweet potato and multiple shoot production per explant, *HortScience* **30**, 1074--1077, 1995.
168. Otani, M., & Shimada, T. Efficient embryogenic callus formation in sweet potato (*Ipomoea batatas* (L.) Lam.), *Breed Sci.* **46**, 257--260, 1996.
169. Al-Mazrooei, S., et al. Optimisation of somatic embryogenesis in fourteen cultivars of sweet potato (*Ipomoea batatas* (L.) Lam.), *Plant Cell Rep.* **16**, 710--714, 1997.
170. Wang, J.S., et al. Efficient embryogenic callus formation and plant regeneration in shoot tip cultures of sweet potato, *Mem. Fac. Agr. Kagoshima Univ.* **34**, 61--64, 1998.
171. Santa-Maria, M., et al. Rapid shoot regeneration in Industrial 'high starch' sweet potato (*Ipomoea batatas* L.) genotypes, *Plant Cell Tiss. Organ Cult.* **97**, 109--117, 2009.

172. Aloufa, M. A. Some factors affecting the callus induction and shoot formation in two cultivars of sweet potato (*Ipomoea batatas* L. POIS), *Cienc. Agrotec.* **26**, 964--969, 2002.
173. Carswell, G.K., & Locy, R.D. Root and shoot initiation by leaf, stem, and storage root explants of sweet potato, *Plant Cell Tiss. Organ Cult.* **3**, 229--236, 1984.
174. Sehgal, C.B. Hormonal control of differentiation in leaf cultures of *Ipomoea batatas* Poir., *Beiträge zur Biologie der Pflanzen* **51**, 47--52, 1975.
175. Chen, L., et al. Approach to establishment of plant regeneration and transformation system in sweet potato (*Ipomoea batatas*) by culture of leaf segments, *Bull. Minamikyushu University* **40 A**, 59--63, 2010.
176. Pido, N., et al. Plant regeneration from leaf discs and stem segments of sweet potato using only NAA as supplementary regulator, *Plant Tiss. Cult. Lett.* **12**, 289--296, 1995.
177. Addae-Frimpomaah, F., et al. The effect of 2, 4-D on callus induction using leaf lobe of sweet potato as a source of explant, *IJAAR* **5** (1), 16--22, 2014a.
178. Ozias-Akins, P. & Perera, S. Regeneration of sweet potato plants from protoplast derived tissues, in *Sweet Potato: Technology for the 21st Century*, W.A. Hill et al, eds., Tuskegee University, Tuskegee, 1992, 61—66.
179. Belarmino, M.M., et al. Plant regeneration from stem and petiole protoplasts of sweet potato (*Ipomoea batatas*) and its wild relative, *I. lacunose*, *Plant Cell Tiss. Organ Cult.* **37**, 145--150, 1994.
180. Liu, J.R., et al. High frequency somatic embryogenesis from cultured shoot apical meristem domes of sweet potato (*Ipomoea batatas*), *SABRAO Journal of Breeding and Genetics* **21**, 93--101, 1989.
181. Sonnino, A., & Mini, P. Somatic embryogenesis in sweet potato *Ipomoea batatas* (L.) Lam, *Acta Hort.* **336**, 239--244, 1993.
182. Litz, R.E., & Conover, R.A. *In vitro* propagation of sweet potato, *HORTSCI*, **13**, 650--660, 1978.
183. Castro, O., & De Andrade, A.G. Meristem culture of sweet potato (*Ipomoea batatas*), *Pesquisa Agropecuária Brasileira Brasília* **30**, 917--922, 1995.
184. Liu, J.R., & Cantliffe, D.J. Somatic embryogenesis and plant regeneration in tissue cultures of sweet potato (*Ipomoea batatas* Poir), *Plant Cell Rep.* **3**, 112--115, 1984.

185. Chee, R.P., & Cantliffe, D.J. Selective enhancement of *Ipomoea batatas* Poir. embryogenic and non-embryogenic callus growth and production of embryos in liquid culture, *Plant Cell Tiss. Organ. Cult.* **15**, 149--159, 1988.
186. Hwang, L.S., et al. Adventitious shoot formation from sections of sweet potato grown *in vitro*, *Sci. Hort.* **20**, 119--129, 1983.
187. Newell, C.A., et al. Transformation of sweet potato (*Ipomoea batatas* (L.) Lam.) with *Agrobacterium tumefaciens* and regeneration of plants expressing cowpea trypsin inhibitor and snowdrop lectin, *Plant Sci.* **107**, 215--227, 1995.
188. Chen, L.Z., et al. Establishment of propagation method in large quantities to produce virus-free plants of sweet potato, *Ipomoea batatas* (L.) Lam. by means of cultures of shoot apex and sucker, *Bull. Fac. Agri, Univ. Miyazaki* **50**, 1--9, 2004 (In Japanese).
189. Murata, T., et al. Callus formation and plant regeneration from petiole protoplast of sweet potato, *Ipomoea batatas* (L.) Lam, *Japan J. Breed.* **37**, 291--298, 1987.
190. Murata, T., et al. Plant regeneration from mesophyll and cell suspension protoplasts of sweet potato, *Ipomoea batatas* (L.) Lam, *Breed. Sci.* **44**, 35--40, 1994.
191. Liu, Q.C., et al. Efficient plant regeneration from embryogenic suspension cultures of sweet potato, *In Vitro Cell Dev. Biol. Plant* **37**, 564--567, 2001.
192. Sultana, R. S., & Rahman, M. M. Cell proliferation and cell aggregate development in suspension culture of sweet potato (*Ipomoea batatas* L.), *Int. J. Biosci.* **1** (6), 6--13, 2011.
193. Feng, C., et al. Cryopreservation of sweetpotato (*Ipomoea batatas*) and its pathogen eradication by cryotherapy, *Biotechnol. Adv.* **29**, 84--93, 2011.
194. González, R.G., et al. Efficient regeneration and *Agrobacterium tumefaciens* mediated transformation of recalcitrant sweet potato (*Ipomoea batatas* L.) cultivars, *Asia Pac. J. Mol. Biol. Biotechnol.* **16**, 25--33, 2008.
195. Chee, R.P., & Cantliffe, D.J. Composition of embryogenic suspension cultures of *Ipomoea batatas* Poir and production of individualized embryos, *Plant Cell Tiss. Organ. Cult.* **17**, 39--52, 1989.
196. Bieniek, M.E., et al. Enhancement of somatic embryogenesis of *Ipomoea batatas* in solid cultures and production of mature somatic embryos in liquid cultures for application to a bioreactor production system, *Plant Cell Tiss. Organ. Cult.* **41**, 1--8, 1995.

197. Liu, Q.C., et al. Cell suspension cultures and efficient plant regeneration in sweet potato, *J. Agr. Biotechnol.* **4**, 238--242, 1996.
198. Liu, Q.C., et al. Establishment of embryogenic cell suspension cultures in sweet potato, *Ipomoea batatas* (L.) Lam, *Acta Agr. Sinica.* **23**, 22--26, 1997.
199. Liu, Q. Sweet potato Omics and Biotechnology in China, *POJ* **4** (6), 295--301, 2011.
200. Xin, S.Y., & Zhang, Z.Z. Explant tissue culture and plantlet regeneration of sweet potato, *Acta Bot. Sinica.* **29**(1), 114--116, 1987.
201. Tan, F., et al. Somatic embryogenesis and plant regeneration in sweet potato. *Acta Agr. Sinica.* **19**, 372--375, 1993.
202. Liu, Q.C., et al. High frequency somatic embryogenesis and plant regeneration in sweet potato, *Ipomoea batatas* (L.) Lam, *J. Agr. Biotechnol.* **1** (1), 84--89, 1993.
203. Gong, Y.F., et al. Advances of *in vitro* culture of sweet potato in China, *Crop Res.* **2**, 46--48, 1998.
204. Gong, Y.F., et al. Effect of NAA and BA on *in vitro* organogenesis of sweet potato, *Journal of Southwest China Normal University* **26**, 443--447, 2001.
205. Gong, Y.F., et al. *In vitro* high frequency direct root and shoot regeneration in sweet potato using the ethylene inhibitor silver nitrate, *S. Afr. J. Bot.* **71** (1), 110--113, 2005.
206. Addae-Frimpomaah, F., et al. Regeneration of three sweet potato (*Ipomea batatas* (L.)) accessions in Ghana via meristem, nodal culture, *Int. J. of Plant breed. Genet.* **8** (3), 121--138, 2014b.
207. Sivparsad, B.J., & Gubba, A. Development of an efficient plant regeneration protocol for sweet potato (*Ipomoea batatas* L.) cv, *Blesbok. Afr. J. Biotechnol.* **11**, 14982--14987, 2012.
208. Perera, S.C., & Ozias-Akins, P. Regeneration from sweet potato protoplasts and assessment of growth conditions for flow-sorting of fusion mixtures, *J AM. SOC. HORTIC. SCI.* **116**, 917--922, 1991.
209. Wu, Y.W., & Ma, C.P. Isolation, culture and callus formation of *Ipomoea batatas* protoplasts, *Acta. Bot. Sinica.* **29** (1), 114--116, 1979.
210. Otani, M., et al. Mesophyll protoplast culture of sweet potato (*Ipomoea batatas* L.), *Plant Sci.* **53**, 157--160, 1987.
211. Sihachakr, D., & Ducreux, G. Plant regeneration from protoplast culture of sweet potato (*Ipomoea batatas* Lam.), *Plant Cell Rep.* **6**, 326--328, 1987.

212. Nishimaki, T., & Nozue, M. Isolation and culture of protoplasts from high anthocyanin-producing callus of sweet potato, *Plant Cell Rep.* **4** (5), 248--251, 1985.
213. Liu, Q.C., et al. Plant regeneration from petiole protoplasts of sweet potato (*Ipomoea batatas* (L.) Lam.) and its related species, *Acta Agr. Sinica.* **21**, 25--28, 1995.
214. Wang, J.S., et al. High frequency plant regeneration from protoplasts of embryogenic callus in sweet potato, *J. Agr. Biotechnol.* **5**, 259--263, 1997.
215. Belarmino, M.M., et al. Shoot formation from protoplast-derived calli of sweet potato and its wild relatives and the initiation of somatic hybrid, *Japan J. Breed.* **43** (Suppl 2), 15--19, 1993.
216. Murata, T., et al. Plant regeneration from fused cells of sweet potato, *Japan J. Breed.* **43** (Suppl 1), 20, 1993.
217. Liu, Q.C., et al. Plant regeneration from *Ipomoea triloba* L. protoplasts, *Japan J. Breed.* **41**, 103--108, 1991.
218. Guo, J.M., et al. Regeneration of plants from *Ipomoea cairica* L. protoplasts and production of somatic hybrids between *I. cairica* L. and sweet potato, *I. batatas* (L.) Lam., *Plant Cell Tiss. Organ. Cult.* **87**, 321--327, 2006.
219. Liu, Q.C., et al. Protoplast fusion and regeneration of interspecific somatic hybrid plants between sweet potato (*Ipomoea batatas* (L.) Lam.) and its related species. *J. Agr. Biotechnol.* **2**, 85--90, 1994.
220. Liu, Q.C., et al. Regeneration and identification of interspecific somatic hybrid plants between sweet potato and *Ipomoea lacunose*, *Acta Agr. Sinica.* **24**, 529--535, 1998.
221. Wang, Y.P., et al. *In vitro* selection and identification of drought-tolerant mutants of sweet potato, *Sci. Agr. Sinica.* **36**, 1000--1005, 2003.
222. Tsai, H.S., & Lin, C.I. The growth of callus induced from *in vitro* culture of sweet potato anthers, *J. Agr. Assoc. China* **81**, 12--19, 1973a.
223. Tsai, H.S., & Lin, C.I. Effects of the compositions of culture media and cultural conditions on growth of callus of sweet potato anther, *J. Agr. Assoc. China* **82**, 310--341, 1973b.
224. Sehgal, C.B. Regeneration of plants from anther culture of sweet potato (*Ipomoea batatas* Poir.), *Zeitschrift Pflanzenphysiol.* **88**, 349--352, 1978.

225. Tsai, H.S., & Tseng, M.T. Embryoid formation and plantlet regeneration from anther callus of sweet potato, *Bot. Bull. Acad. SINICA* **20**, 117--122, 1979.
226. Nishiguchi, M., et al. Stable transformation of sweet potato by electroporation, *In Vitro Cell Dev. Plant* **28**, 126, 1992.
227. García, R., et al. Sweet potato (*Ipomoea batatas* L.) regeneration and transformation technology to provide weevil (*Cylas formicarius*) resistance. Field trial results. In: Arencibia AD (eds) Plant Genetic Engineering: Towards the Third Millennium. *Elsevier Science B.V.* 112--117, 2000.
228. Prakash, C.S., & Varadarajan, U. Genetic transformation of sweet potato by particle bombardment, *Plant Cell Rep.* **11**, 53--57, 1992.
229. Yang, K.Y., et al. Production of transgenic sweet potato (*Ipomoea batatas* (L.) Lam.) lines via microprojectile bombardment, *Korean J. Breed.* **37**, 236--240, 2005.
230. Fromm, M., et al. Expression of genes transferred into monocot and dicot plant cells by electroporation, *Proc. Natl. Acad. Sci.* **82**, 5824--5828, 1985.
231. Dhir, S.K., et al. Factors affecting transient gene expression in electroporated soybean (*Glycine max* L.) protoplasts, *Plant Cell Rep.* **10**, 106--110, 1991.
232. Joersbo, M., & Brunstedt, J. Electroporation: mechanism and transient expression, stable transformation and biological effects in plant protoplasts, *Physiol. Plant.* **81** (2), 256--264, 1991.
233. Mitchell, T.D., et al. Electroporation mediate transient gene expression in intact cell of sweet potato, *In Vitro Cell Dev. Plant.* **34**, 319--324, 1998.
234. Lawton, R., et al. Expression of green fluorescent protein genes in sweet potato tissues, *Plant Mol. Biol. Rep.* **18** (2), 139, 2000.
235. Winfield, S., et al. Transformation of sweet potato tissues with green fluorescent protein gene, *In Vitro Cell Dev. Plant.* **37**, 648--653, 2001.
236. Okada, Y., et al. Virus resistance in transgenic sweet potato (*Ipomoea batatas* L. (Lam)) expressing the coat protein gene of sweet potato feathery mottle virus, *Theor. Appl. Genet.* **103**, 743--751, 2001.
237. Babu, R.M., et al. Advances in genetically engineered (transgenic) plants in pest management— An overview, *Crop Prot.* **22**, 1071--1086, 2003.

238. Yi, G., et al. Production of herbicide-resistant sweet potato plants transformed with the *bar* gene, *Biotechnol. Lett.* **29**, 669--675, 2007.
239. Lim, S., et al. Enhanced tolerance to transgenic sweet potato plants that express both CuZnSOD and APX in chloroplasts to methyl viologen mediated oxidative stress and chilling, *Mol. Breeding* **19** (30), 227--239, 2007.
240. Dodds, J.H., Merzdorf, C., Zambrano, V. & Siguenas, C. Potential use of *Agrobacterium*-mediated gene transfer to confer insect resistance in sweet potato, in *Sweet potato pest management: a global perspective*, R.K. Jansson et al, eds., West View Press, Oxford, U.K., 1991, 203— 219,
241. Otani, M., et al. Transformation of sweet potato (*Ipomoea batatas* (L.) Lam.), *Plant Sci.* **94**, 151--159, 1993.
242. Carelli, M.L.D., Skirvin, R.M. & Harry, D.E. Transformation and regeneration studies of “Jewel” sweet potato, in *Sweet Potato Technology for the 21st Century*, W.A. Hill et al, eds., Tuskegee University, Tuskegee. 52-60, 1991.
243. Prakash, C.S. & Varadarajan, U. Expression of foreign genes in transgenic sweet potatoes, in Proceedings of the International Society and Plant Molecular Biology, Tucson, AZ, USA, October 8-12, 1991.
244. Otani, M., et al. Genetic transformation of sweet potato (*Ipomoea batatas* (L.) Lam.) by *Agrobacterium tumefaciens*, *Acta Hort. (ISHS)* **560**, 193--196, 2001.
245. Wakita, Y., et al. A tobacco microsomal ω -3 fatty acid desaturase gene increases the linolenic acid content in transgenic sweet potato (*Ipomoea batatas*), *Plant Cell Rep.* **20**, 244--249, 2001.
246. Kimura, T., et al. Absence of amylose in sweet potato [*Ipomoea batatas* (L.) Lam.] following the introduction of granule-bound starch synthase I cDNA, *Plant Cell Rep.* **20**, 663--666, 2001.
247. Otani, M., et al. Production of herbicide-resistant sweet potato (*Ipomoea batatas* (L.) Lam.) plants by *Agrobacterium tumefaciens*-mediated transformation, *Breed. Sci.* **53**, 145--148, 2003.
248. Song, G.Q., et al. Efficient *Agrobacterium tumefaciens* mediated transformation of sweet potato (*Ipomoea Batatas* (L.) Lam) from stem explants using a two-step kanamycin-hygromycin selection method, *In Vitro Cell Dev. Plant* **40**, 359--365, 2004.

249. Shimada, T., et al. Increase of amylose content of sweet potato starch by RNA interference of the starch branching enzyme II gene (*IbSBE II*), *Plant Biotechnol.* **23**, 85--90, 2006.
250. Luo, H.R., et al. Rapid genetic transformation of sweet potato (*Ipomoea batatas* (L.) Lam.) via organogenesis, *Afr. J. Biotechnol.* **5**, 1851--1857, 2006.
251. Gama, M.I.C., et al. Transgenic sweetpotato plants obtained by *Agrobacterium tumefaciens* mediated transformation, *Plant Cell Tiss. Org. Cult.* **46**, 237--244, 1996.
252. Otani, M., et al. Transgenic plant production from embryogenic callus of sweet potato (*Ipomoea batatas* (L.) Lam.) using *Agrobacterium tumefaciens*, *Plant Biotechnol.* **15**, 11--16, 1998.
253. Chen, L., et al. Establishment of *Agrobacterium*-Mediated Transformation System in Sweet Potato (*Ipomoea batatas*) by Culture of Leaf Segments for Functional Analysis of ASG-1, an Apomixis-Specific Gene, *British Biotechnology Journal*, ISSN, **3** (4), 2231--2927, 2013.
254. Yu, B., et al. Efficient *Agrobacterium tumefaciens* mediated transformation using embryogenic suspension cultures in sweet potato, *Ipomoea batatas* (L.) Lam, *Plant Cell Tiss. Organ. Cult.* **90**, 265--273, 2007.
255. Zang, N., et al. Efficient production of transgenic plants using the *bar* gene for herbicide resistance in sweet potato, *Sci Horti* 122, 649--653, 2009.
256. Gao, S., et al. Enhanced stem nematode resistance of transgenic sweet potato plants expressing oryzacystatin-I gene, *Agr. Sci.China* **10**, 519--525, 2011.
257. Yang, J., et al. Efficient embryogenic suspension culturing and rapid transformation of a range of elite genotypes of sweet potato (*Ipomoea batatas* [L.] Lam.), *Plant Sci.* **181**, 701--711, 2011.
258. Zhai, H., & Liu, Q.C. Studies on the genetic transformation of embryogenic suspension cultures in sweet potato, *Sci. Agr. Sinica* **36**, 487--491, 2003.
259. Jiang, S.J., et al. Regeneration of sweet potato transgenic plants with *oryzacystatin-I* (*OC I*) gene, *J. Agr. Biotechnol.* **12**, 34--37, 2004b.
260. Moran, R., et al. Transgenic sweet potato plants carrying the delta-endotoxin gene from *Bacillus thuringiensis* var. tenebrionis, *Plant Sci.* **139**, 175--184, 1998.

261. Gichuki, S.T., et al. Development of virus resistance sweet potato using biotechnological approaches in Kenya, In: *Proceeding of Symposium International Society of Tropical Root Crops*, held in Arusha, Tanzania, 9-14 November 2003, ISRTC Publications, Croydon, England, UK, 124—128, 2003.
262. Haque, A.K.M., et al. Analysis of transitive RNA silencing after grafting in transgenic plants with the coat protein gene of *Sweet potato feathery mottle virus*, *Plant Mol. Biol.* **63**, 35--47, 2007.
263. Okada, Y., & Saito, A. Evaluation of resistance to complex infection of SPFMVs in transgenic sweet potato, *Breeding Sci.* **58**, 243--250, 2008.
264. Kreuze, J.F., et al. RNA silencing-mediated resistance to a crinivirus (Closteroviridae) in cultivated sweet potato (*Ipomoea batatas* L.) and development of sweet potato virus disease following co-infection with a potyvirus, *Mol. Plant Pathol.* **9**, 589--598, 2008.
265. Sivparsad, B.J., & Gubba, A. Development of transgenic sweet potato with multiple virus resistance in South Africa (SA), *Transgenic Res.* **23**, 377--388, 2014.
266. Mohan, C., & Nair, A. G. H. Characterization of Genes and Promoters, Transformation and Transgenic Development in Sweet Potato, in Nedunchezhiyan M, Byju G (Eds) Sweet Potato. *Fruit, Vegetable and Cereal Science and Biotechnology*, **6** (Special Issue 1), 43—56, 2012.
267. Muramoto, N., et al. Transgenic sweet potato expressing thionin from barley gives resistance to black rot disease caused by *Ceratocystis fimbriata* in leaves and storage roots, *Plant Cell Rep.* **31**, 987--997, 2012.
268. Lu, Y.Y., et al. Over expression of CuZn superoxide dismutase (*CuZn SOD*) ascorbate peroxidase (*APX*) in transgenic sweet potato enhances tolerance and recovery from drought stress, *Afr. J. Biotechnol.* **9**, 8378--8391, 2010.
269. Wang, X., et al. Studies on salt tolerance of transgenic sweet potato which harbours two genes expressing CuZn superoxide dismutase and ascorbate peroxidase with the stress-inducible SWPA2 promoter, *Plant Gene and Trait* **3**, 6--12, 2012.
270. Park, S.C., et al. Sweet potato late embryogenesis abundant 14 (*IbLEA14*) gene influences lignifications and increases osmotic- and salt stress-tolerance of transgenic calli, *Planta* **233**, 621--634, 2011.

-
271. Kim, Y.H., et al. *SCOF-1*-expressing transgenic sweet potato plants show enhanced tolerance to low-temperature stress, *Plant Physiol. Bioch.* **49**, 1436--1441, 2011.
272. Kim, S.H., et al. Down-regulation of β - carotene hydroxylase increases β -carotene and total carotenoids enhancing salt stress tolerance in transgenic cultured cells of sweet potato, *Phytochemistry* **74**, 69--78, 2012.
273. Kim, S.H., et al. Cloning and characterization of an *Orange* gene that increases carotenoid accumulation and salt stress tolerance in transgenic sweetpotato cultures, *Plant Physiol. Bioch.* **70**, 445--454, 2013a.
274. Kim, S.H., et al. Downregulation of the lycopene ϵ -cyclase gene increases carotenoid synthesis via the β -branch-specific pathway and enhances salt-stress tolerance in sweetpotato transgenic calli, *Physiol. Plant* **147**, 432--442, 2013b.
275. Yi, G., e. al. Production of herbicide-resistant sweet potato plants transformed with the bar gene, *Biotechnol. Lett.* **29**, 669--675, 2007.
276. Choi, H.J., et al. Production of herbicide-resistant transgenic sweet potato plants through *Agrobacterium tumefaciens* method, *Plant Cell Tiss. Organ Cult.* **91**, 235--242, 2007.
277. Anwar, N., et al. Transgenic sweet potato expressing mammalian cytochrome P450, *Plant Cell Tiss. Organ Cult.* **105**, 219--231, 2011.
278. Otani, M., et al. Inhibition of the gene expression for granule-bound starch synthase I by RNA interference in sweet potato plants, *Plant Cell Rep.* **26**, 1801--1807, 2007.
279. Takahata, Y., et al. Inhibition of the expression of the starch synthase II gene leads to lower pasting temperature in sweet potato starch, *Plant Cell Rep.* **29**, 535--543, 2010.
280. Tanaka, M., et al. Altered carbohydrate metabolism in the storage roots of sweet potato plants overexpressing the SRF1 gene, which encodes a Dof zinc finger transcription factor, *Planta* **230**, 737--746, 2009.
281. Santa-Maria, M.C., et al. Starch self-processing in transgenic sweet potato roots expressing a hyperthermophilic β -amylase, *Biotechnol. Progr.* **27**, 351--359, 2011.
282. Kroger, M. Nutritionally Improved Sweetpotato. *Compr. Rev. Food Sci. F.* **7** (1), 81--91, 2008.
283. Toyama, J., et al. Selection of sweetpotato lines with high protein content and/or low trypsin inhibitor activity, *Breeding Sci.* **56**, 17--23, 2006.

-
284. Berberich, T., et al. Production of mouse adiponectin, an anti-diabetic protein, in transgenic sweet potato plants, *J. Plant Physiol.* **162**, 1169--1176, 2005.
285. Kim, T.G., et al. Expression of nutritionally well-balanced protein, AmA1, in *Saccharomyces cerevisiae*, *Biotechnol. Bioprocess Eng.* **6**, 173--178, 2001.
286. Chakraborty, S., et al. Premature truncation of RNA polymerase II mediated transcription of a seed protein in *Schizosaccharomyces pombe*, *Nucleic Acids Res.* **30**, 2940--2949, 2002.
287. Sarmah, B. et al. Plant pre-mRNA splicing in fission yeast, *Schizosaccharomyces pombe*, *Biochem. Biophys. Res. Commn.* **293**, 1209--1216, 2002.
288. Tamás, C., et al. Transgenic approach to improve wheat (*Triticum aestivum* L.) nutritional quality, *Plant Cell Rep* **28**, 1085--1094, 2009.
289. Zolla, L., et al. Proteomics as a complementary tool for identifying unintended side effects occurring in transgenic maize seeds as a result of genetic modifications, *J. Proteome Res.* **7**, 1850--1861, 2008.
290. Gong, C. Y., & Wang, T. Proteomic evaluation of genetically modified crops: current status and challenges, *Front. Plant Sci.* **4**, 41, 2013.
291. Coll, A., et al. Lack of repeatable differential expression patterns between MON810 and comparable commercial varieties of maize, *Plant Mol Biol.* **68** (1-2), 105--117, 2008.
292. Coll, A., et al. Gene expression profiles of MON810 and comparable non-GM maize varieties cultured in the field are more similar than are those of conventional lines, *Transgenic Res.* **18** (5), 801--808, 2009.
293. Kogel, K.H., et al. Transcriptome and metabolome profiling of field-grown transgenic barley lack induced differences but show cultivar-specific variances, *Proc. Natl. Acad. Sci.* **107** (14), 6198--6203, 2010.
294. Montero, M., et al. Only half the transcriptomic differences between resistant genetically modified and conventional rice are associated with the transgene. *Plant Biotechnol. J.* **9**, 693--702, 2011.
295. Ruebelt, M. C. et al. Application of two-dimensional gel electrophoresis interrogate alterations in the proteome of genetically crops. 1. Assessing unintended effects, *J. Agric. Food Chem.* **54** (6), 2154- 61, 2006.

296. Corpillo, D., et al. Proteomics as a tool to improve investigation of substantial equivalence in genetically modified organisms: the case of a virus-resistant tomato, *Proteomics* **4**,193--200, 2004.
297. Di Carli, M., et al. Leaf proteome analysis of transgenic plants expressing antiviral antibodies, *J. Proteome Res.* **8**, 838--848, 2009.
298. Chakraborty, N., et al. Reduction of oxalate levels in tomato fruit and consequent metabolic remodeling following overexpression of a fungal oxalate decarboxylase, *Plant Physiol.* **162**, 364--378, 2013.
299. Lehesranta, S. J., et al. Comparison of tuber proteomes of potato varieties, landraces and genetically modified lines, *Plant Physiol.* **138**, 1690--1699, 2005.
300. Islam, N., et al. Decreased accumulation of glutelin types in rice grains constitutively expressing a sunflower seed albumin gene, *Phytochemistry* **66**, 2534--2539, 2005.
301. Gong, C. Y., et al. Proteomics insight into the biological safety of transgenic modification of rice as compared with conventional genetic breeding and spontaneous genotypic variation, *J. Proteome Res.* **11**, 3019--3029, 2012.
302. Barros, E., et al. Comparison of two GM maize varieties with a near-isogenic non-GM variety using transcriptomics, proteomics and metabolomics, *Plant Biotechnol. J.* **8**, 436--451, 2010.
303. Coll, A., et al. Proteomic analysis of MON810 and comparable non-GM maize varieties grown in agricultural fields, *Transgenic Res.* **20**, 939--949, 2011.
304. Balsamo, G. M., et al. Proteomic analysis of four Brazilian MON810 maize varieties and their four non-genetically-modified isogenic varieties, *J. Agric. Food Chem.* **59**, 11553--11559, 2011.
305. Brandao, A. R., et al. Image analysis of two-dimensional gel electrophoresis for comparative proteomics of transgenic and non-transgenic soybean seeds, *J. Proteomics* **73**, 1433--1440, 2010.
306. Barbosa, H. S., et al. New insights on proteomics of transgenic soybean seeds: evaluation of differential expressions of enzymes and proteins, *Anal. Bioanal. Chem.* **402**, 299--314, 2012.
307. Rocco, M., et al. The expression of tomato prosystemin gene in tobacco plants highly affects host proteomic repertoire, *J. Proteomics* **71**, 176--185, 2008.

308. Di Carli, M., et al. Proteomic analysis of the plant-virus interaction in cucumber mosaic virus (CMV) resistant transgenic tomato, *J. Proteome Res.* **9**, 5684--5697, 2010.
309. Di Luccia, A., et al. A proteomic approach to study protein variation in GM durum wheat in relation to technological properties of semolina, *Ann. Chim.* **95**, 405--414, 2005.
310. Scossa, F., et al. Comparative proteomic and transcriptional profiling of a bread wheat cultivar and its derived transgenic line overexpressing a low molecular weight glutenin subunit gene in the endosperm, *Proteomics* **8**, 2948--2966, 2008.
311. Clarke, J.D., et al. Assessment of Genetically Modified Soybean in Relation to Natural Variation in the Soybean Seed Metabolome, *Scientific Rep.* **3**, 3082, 2013.
312. Catchpole, G.S., et al. Hierarchical metabolomics demonstrates substantial compositional similarity between genetically modified and conventional potato crops, *Proc. Natl. Acad. Sci.* **102**, 14458--14462, 2005.
313. Chassy, B. M. Can -omics inform a food safety assessment? *Regul. Toxicol. Pharmacol.* **58**, S62--S70, 2010.
314. Szopa, J. Transgenic 14-3-3 isoforms in plants: the metabolite profiling of repressed 14-3-3 protein synthesis in transgenic potato plants, *Biochem. Soc. Trans.* **30**, 405--410, 2002.
315. Roessner, U., et al. Metabolic profiling allows comprehensive phenotyping of genetically or environmentally modified plant systems, *Plant Cell* **13**, 11--29, 2001.
316. Baker, J. M., et al. A metabolomic study of substantial equivalence of field-grown genetically modified wheat, *Plant Biotechnol. J.* **4**, 381--392, 2006.
317. Mattoo, A. K., et al. Nuclear magnetic resonance spectroscopy-based metabolite profiling of transgenic tomato fruit engineered to accumulate spermidine and spermine reveals enhanced anabolic and nitrogen-carbon interactions, *Plant Physiol.* **142**, 1759--1770, 2006.
318. Komatsu, S., et al. Application of proteomics for improving crop protection/artificial regulation, *Front. Plant Sci.* **4**, 1--3, 2013.
319. Ghazi, A., et al. Protein isolates from egyptian sweet potato leaves, *Die Nahrung* **33** (2), 145--151, 1989.
320. Jiang, Y., et al. A proteomic analysis of storage stress responses in *Ipomoea batatas* (L.) Lam. tuberous root, *Mol. Biol. Rep.* **39**, 8015--8025, 2012.
321. Jiang, Y.S., et al. Characterization and expression of Rubisco activase genes of *Ipomoea batatas*, *Mol. Biol. Rep.* **40** (11), 6309--6321, 2013.

322. Lee, J.J., et al. Comparative proteomic study between tuberous roots of light orange- and purple-fleshed sweetpotato cultivars, *Plant Sci.* **193-194**, 120--129, 2012.
323. Mukherjee, A. Tuber Crops, in *Biotechnology and its Application in Horticulture*, S.P. Ghosh eds., Narosa Publishing House, New Delhi, 1999, 267-294.
324. Sambrook, J. & Russell, W. R. *Molecular Cloning: A Laboratory Manual*, 3rd ed., Cold Spring Harbor Laboratory Press, NY, 2001.
325. Ditta, G., et al. Broad host-range DNA colony system for gram-negative bacteria- construction of a gene bank of rhizobium malilotis, *Proc. Natl. Acad. Sci.* **77**, 7347--7351, 1980.
326. Van Haute, E., et al. Intergeneric transfer and exchange recombination of restriction fragments cloned in pBR322 a novel strategy for reversed genetics of the Ti plasmid of *Agrobacterium tumefaciens*, *EMBO J.* **2**, 411--418, 1983.
327. Alam, I., et al. Effect of Growth Regulators on Meristem Culture and Plantlet Establishment in Sweet Potato [*Ipomoea Batatas*' (L.) Lam.], *Plant Omics.* **3**, 35--39, 2010.
328. Fior, S., & Gerola, P.D. Impact of ubiquitous inhibitors on the *GUS* gene reporter system: evidence from the model plants *Arabidopsis*, tobacco and rice and correction methods for quantitative assays of transgenic and endogenous *GUS*, *Plant Methods* **5**, 19, 2009.
329. Lambardi, M., Ozudogru, E. A. & Benelli, C. Cryopreservation of embryogenic cultures, in *Plant cryopreservation—a practical guide*, B. M. Reed eds., Springer Science and Business Media, New York, 2008, 177—210
330. Zhang, D., Cipriani, G., Rety, I., Golmirzaie, A., Smit, S. & Michaud, D. Expression of protease inhibitors in sweet potato, in *Recombinant Protease Inhibitors in Plants*, D. Michaud eds., Landes Bioscience, USA, 2000,167—178.
331. Zhang, L., et al. An efficient wheat transformation procedure: transformed calli with long-term morphogenic potential for plant regeneration, *Plant Cell Rep.* **19**, 241--250, 2000.
332. Bhat, S.R., & Srinivasan, S. Molecular and genetic analyses of transgenic plants: considerations and approaches, *Plant Sci.* **163**, 673--681, 2002.
333. Pawlowski, W.P., & Somers, D.A. Transgenic inheritance in plants genetically engineered using microprojectile bombardment, *Mol. Biotechnol.* **6**, 17--30, 1996.

334. Tizaoui, K., & Kchouk, M.E. Genetic approaches for studying inheritance and genetic recombination in three successive generations of transformed tobacco, *Genet. Mol. Biol.* **35**, 640--649, 2012.
335. Somers, D.A., & Makarevitch, I. Transgene integration in plants poking or patching holes in promiscuous genomes? *Curr. Opin. Biotechnol.* **15**, 126--131, 2004.
336. De Neve, N. et al. T-DNA integration patterns in cotrans formed plant cells suggest that T-DNA repeats originate from cointegration of separate T-DNAs, *Plant J.* **11**, 15--29, 1997.
337. Takano, M., et al. The structures of integration sites in transgenic rice: Three structures of integration sites in transgenic rice, *Plant J.* **11**, 353--361, 1997.
338. Topping, J.F., et al. Functional tagging of regulatory elements in the plant genome, *Development* **112**, 1009--1019, 1991.
339. Dellaporta, S.L., et al. A plant DNA miniprep: version II, *Plant Mol. Biol. Rept.* **1**, 19--21, 1983.
340. Mason, G., et al. Estimating the number of integrations in transformed plants by quantitative real-time PCR, *BMC Biotechnol.* **2**, 20, 2002.
341. Pandey, V., et al. Standardization of qualitative and quantitative polymerase chain reaction methods in transgenic *Indica* rice, *IJBAS* **2**(4) 356--365, 2013.
342. Jefferson, R.A. Assaying chimeric genes in plants: the GUS gene fusion system, *Plant Mol. Biol. Rep.* **5**, 387--405, 1987.
343. Hoffman, L. M., et al. A modified storage protein is synthesized, processed, and degraded in the seeds of transgenic plants, *Plant Mol. Biol.* **11** (6), 717--729, 1988.
344. Laemmli, U.K. Cleavage of structural proteins during the assembly of the head of the bacteriophage T4, *Nature* **227**, 680--685, 1970.
345. Kim, Y.H., et al. Molecular characterization of a *cDNA* encoding DRE-binding transcription factor from dehydration-treated fibrous roots of sweet potato, *Plant Physiol. Biochem.* **46**, 196--204, 2008.
346. Outchkourov, N.S., et al. Expression of sea anemone equistatin in potato. Effects of plant proteases on heterologous protein production, *Plant Physiol.* **133**, 379--390, 2003.
347. Raeymaekers, L. Basic principles of quantitative PCR, *Mol. Biotechnol.* **15** (2), 115--122, 2000.

348. Ingham, D.J., et al. Quantitative real-time PCR assay for determining transgene copy number in transformed plants, *Biotechniques* **31** (1), 132--140, 2001.
349. Pfaffl, M.W. Quantification strategies in real-time PCR, in *A–Z of quantitative PCR*, S.A. Bustin, eds., International University Line (IUL), CA, 2004, 87—112
350. Park, S.C., et al. Stable Internal Reference Genes for the Normalization of Real-Time PCR in Different Sweetpotato Cultivars Subjected to Abiotic Stress Conditions, *Plos ONE* **7**, e51502, 2012.
351. Huggett, J., et al. Real-time RT–PCR normalisation: strategies and considerations, *Genes Immun.* **6**, 279--284, 2005.
352. Gutierrez, L., et al. The lack of a systemic validation of reference genes: serious pitfall undervalued in reverse transcription–polymerase chain reaction (RT–PCR) analysis in plants, *Plant Biotechnol. J.* **6**, 609--618, 2008.
353. Gue´nin, S., et al. Normalization of qRT–PCR data: the necessity of adopting a systematic, experimental conditions-specific, validation of references, *J. Exp. Bot.* **60**, 487-493, 2009.
354. Dekkers, B.J.W., et al. Identification of reference genes for RT–qPCR expression analysis in Arabidopsis and tomato seeds, *Plant cell physiol.* **53** (1), 28--37, 2012.
355. Remans, T., et al. Normalisation of real-time RT-PCR gene expression measurements in *Arabidopsis thaliana* exposed to increased metal concentrations, *Planta* **227**, 1343--1349, 2008.
356. Schmidt, G., & Delaney, S. Stable internal reference genes for normalization of real-time RT-PCR in tobacco (*Nicotiana tabacum*) during development and abiotic stress, *Mol. Genet. Genomics* **283**, 233--241, 2010.
357. Løvdal, T., & Lillo, C. Reference gene selection for quantitative real-time PCR normalization in tomato subjected to nitrogen, cold, and light stress, *Anal. Biochem.* **387**, 238--242, 2009.
358. Nicot, N., et al. Housekeeping gene selection for real-time RT-PCR normalization in potato during biotic and abiotic stress, *J. Exp. Bot.* **56**, 2907--2914, 2005.
359. Xu, M., et al. Reference gene selection for quantitative real-time polymerase chain reaction in Populus, *Anal. Biochem.* **408**, 337--339, 2011.

360. Kong, Q., et al. Screening Suitable Reference Genes for Normalization in Reverse Transcription Quantitative Real-Time PCR Analysis in Melon, *PloS One* **9** (1), e87197, 2014.
361. Huang, S., et al. Transgenic studies on the involvement of cytokinin and gibberellin in male development, *Plant. Physiol.* **131**, 1270--1282, 2003.
362. Veluthambi, K., et al. The current status of plant transformation technologies, *Curr. Sci.* **84**, 368--380, 2003.
363. Hahn, S.K. & Hozyo, Y. Sweet Potato, in *the Physiology of Tropical Field Crops*, P.R. Goldsworthy et al, eds., John Wiley and Sons Ltd., London, 1984, 551— 567,
364. Wilson, J.W. Control of crop processes, in *Crop Processes in Controlled Environments*, A.R. Rees et al, eds., Academic Press, London and New York, 1972, 7— 30.
365. Riesmeier, J. W., et al. Evidence for an essential role of the sucrose transporter in phloem loading and assimilate partitioning, *EMBO J.* **13**, 1--7, 1994.
366. Jonik, C., et al. Simultaneous boosting of source and sink capacities doubles tuber starch yield of potato plants, *Plant Biotechnol. J.* **10**, 1088--1098, 2012.
367. Schubert, D., et al. Silencing in *Arabidopsis* T-DNA transformants: the predominant role of a gene-specific RNA sensing mechanism versus position effects, *Plant Cell* **16**, 2561--2572, 2004.
368. Butaye, K. M., et al. Approaches to minimize variation of transgene expression in plants, *Mol. Breeding* **16** (1), 79--91, 2005.
369. Oltmanns, H., et al. Generation of backbone-free, low transgene copy plants by launching T-DNA from the *Agrobacterium* chromosome, *Plant Physiol.* **152**, 1158--1166, 2010.
370. Gahakwa, D., et al. Transgenic rice as a system to study the stability of transgene expression: multiple heterologous transgenes show similar behavior in diverse genetic backgrounds, *Theor. Appl. Genet.* **101**, 388--399, 2000.
371. Hadi, F., et al. Development of quantitative competitive PCR for determination of copy number and expression level of the synthetic glyphosate oxidoreductase gene in transgenic canola plants, *Electron J. Biotechn.* **15** (4), 2--14, 2012.
372. Hobbs, S.L.A., et al. Transgene copy number can be positively or negatively associated with transgene expression, *Plant Mol. Biol.* **21**, 17--26, 1993.

373. Pinheiro, T. T., et al. Early-flowering sweet orange mutant 'x11' as a model for functional genomic studies of Citrus, *BMC Res. Notes* **7** (1), 511, 2014.
374. Agrawal, L., et al. Comparative proteomics of tuber induction, development and maturation reveal the complexity of tuberization process in potato (*Solanum tuberosum* L.), *J. Proteome Res.*, **7**, 3803--3817, 2008.
375. Schmidt, M.A., & Herman, E.M. Proteome rebalancing in soybean seeds can be exploited to enhance foreign protein accumulation, *Plant Biotechnol. J.* **6**, 832--842, 2008.
376. Schmidt, M. A., et al. Silencing of soybean seed storage proteins results in rebalanced protein composition preserving seed protein content without major collateral changes in the metabolome and transcriptome, *Plant Physiol.* **156**, 330--345, 2011.
377. Oszvald, M., et al. Wheat storage proteins in transgenic rice endosperm, *J. Agric. Food Chem.* **61**, 7606--7614, 2013.
378. Wu, Y., & Messing, J. Proteome balancing of the maize seed for higher nutritional value, *Front. Plant Sci.* **5**, 240, 2014.
379. Moutot, F., et al. Relationship between photosynthesis and protein synthesis in maize. I. Kinetics of translocation of the photoassimilated carbon from the ear leaf to the seed, *Plant Physiol.* **80**, 211--215, 1986.
380. Rommens, C. M., et al. The intragenic approach as a new extension to traditional plant breeding, *Trends Plant Sci.* **12**, 397--403, 2007.
381. Purrington, C.B., & Bergelson, J. Assessing weediness of transgenic crops: industry plays plant ecologist, *Tree* **10**, 340--342, 1995.
382. Huang, A.S., et al. Content of alpha-, beta-Carotene, and dietary fiber in 18 sweetpotato varieties grown in Hawaii, *J. Food Comp. Anal.* **12**, 147--151, 1999.
383. Kennedy, G., & Burlingame, B. Analysis of food composition data on rice from a plant genetic resources perspective, *Food Chem.* **80**, 589--596, 2003.
384. Zeller, S.L., et al. Transgene x environment interactions in genetically modified wheat, *PLoS ONE* **5**, e11405, 2010.
385. Ssemakula, G., et al. Stability of total carotenoid concentration and fresh yield of selected yellow-fleshed cassava (*Manihot esculenta* Crantz), *Journal of Tropical Agriculture* **45**, (1-2), 14--20, 2007.

386. Maloof, J. N., et al. Leaf J: an image plugin for semi-automated leaf shape measurement, *J. Vis. Exp.* **71**, e50028, 2013.
387. Association of official analytical chemists (AOAC). *Official methods of analysis*, 17th ed., Association of official analytical chemists, Washington, D.C. 2000.
388. Sadasivam, S. & Manickam, A. *Biochemical Methods for Agricultural Sciences*, Wiley Eastern Limited, New Delhi, 1992.
389. Roessner, U., et al. Technical advance: simultaneous analysis of metabolites in potato tuber by gas chromatography-mass spectrometry, *Plant J.* **23**, 131--142, 2000.
390. Roessner, U., et al. High-resolution metabolic phenotyping of genetically and environmentally diverse potato tuber systems. Identification of phenocopies, *Plant Physiol.* **127**, 749--764, 2001.
391. Robertson, J.A., et al. Hydration properties of dietary fibre and resistant starch: a European collaborative study, *Lebensm.-Wiss. u.-Technol.*, **33**, 72--79, 2000.
392. Horsley, S. B., & Gottschalk, K. W. Leaf area and net photosynthesis during development of *Prunus serotina* seedlings, *Tree Physiol.* **12**, 55--69, 1993.
393. Harijono T. E., et al. Effect of Blanching on Properties of Water Yam (*Dioscorea alata*) Flour. *Adv. J. Food Sci. Technol.* **5**, 1342--1350, 2013.
394. Traynham, T. L., et al. Evaluation of water-holding capacity for wheat-soy flour blends, *J. Am. Oil Chem. Soc.* **84**, 151--155, 2007.
395. Sweetlove, L.J., et al. The control of source to sink carbon flux during tuber development in potato, *Plant J.* **15**, 697--706, 1998.
396. Moutot, F., et al. Relationship between photosynthesis and protein synthesis in maize. I. Kinetics of translocation of the photoassimilated carbon from the ear leaf to the seed, *Plant Physiol.* **80**, 211--215, 1986.
397. Zhu, X.G., et al. Improving photosynthetic efficiency for greater yield, *Annu. Rev. Plant Biol.* **61**, 235--261, 2010.
398. Altenbach, S.B., et al. Enhancement of the methionine content of seed proteins by the expression of a chimeric gene encoding a methionine-rich protein in transgenic plants, *Plant Mol. Biol.* **13**, 513--522, 1989.

399. Zheng, Z., et al. The bean seed storage protein [beta] - phaseolin is synthesized, processed, and accumulated in the vacuolar type-II protein bodies of transgenic rice endosperm. *Plant Physiol.* **109**, 777--786, 1995.
400. Falco, S.C., et al. Transgenic canola and soybean seeds with increased lysine. *Biotechnology (N Y)*, **13**, 577--582, 1995.
401. Collins, W.W. & Walter, W.M. Jr. Fresh Roots for Human Consumption, in *Sweet Potato products: A Natural Resource for the Tropics*, J. C. Bouwkamp, eds. CRC Press, Florida, 1985, 176-200.
402. Azevedo, A.M., et al. Influence of harvest time and cultivation sites on the productivity and quality of sweet potato, *Hortic. Bras.* **32**, 21--27, 2014.
403. Druege, U, et al. Nitrogen- and storage-affected carbohydrate partitioning in high-light-adapted *Pelargonium* cuttings in relation to survival and adventitious root formation under low light, *Ann. Bot. (Lond.)* **94**, 831--842, 2004.
404. Gibbon, B.C., et al. Altered starch structure is associated with endosperm modification in Quality Protein Maize, *Proc. Natl. Acad. Sci.* **100**(26), 15329--15334, 2003.
405. Kizil, R., et al. Characterization of irradiated starches by using FT-Raman and FTIR spectroscopy, *J. Agric. Food Chem.* **50**, 3912--3918, 2002.
406. Keymanesh, K., et al. Metabolome comparison of transgenic and non-transgenic rice by statistical analysis of FTIR and NMR spectra, *Rice Sci.* **16**, 119--123, 2009.
407. Dunn, W. B., & Ellis, D. I. Metabolomics: current analytical platforms and methodologies, *TrAC Trend Anal. Chem.* **24**(4), 285--294, 2005.
408. Trethewey, R.N., et al. Metabolic profiling: a Rosetta stone for genomics? *Curr. Opin. Plant Biol.* **2**, 83--85, 1999.
409. Choi, H. K., et al. Metabolic fingerprinting of wild type and transgenic tobacco plants by ¹H NMR and multivariate analysis technique, *Phytochemistry* **65**(7), 857--864, 2004.
410. Roessner-Tunali, U., et al. Metabolic profiling of transgenic tomato plants overexpressing hexokinase reveals that the influence of hexose phosphorylation diminishes during fruit development, *Plant Physiol.* **133**, 84--99, 2003.
411. Rochfort, S. Metabolomics reviewed: a new "omics" platform technology for systems biology and implications for natural products research, *J. Nat Products* **68**(12), 1813--1820, 2005.

412. Kim, D.O., et al. Antioxidant capacity of phenolic phytochemicals from various cultivars of plums, *Food Chem.* **81**, 321--326, 2003.
413. Mancinelli, A., et al. Anthocyanin production in chl-rich and chl-poor seedlings, *Plant Physiol.* **86**, 652--654, 1988.
414. Koala, M., et al. Evaluation of eight orange fleshed sweet potato (OFSP) varieties for their total antioxidant, total carotenoid and polyphenolic contents, *Journal of Natural Sciences Research* **3**, 67--72.
415. Hagenimana, V., et al. Carotenoid contents in fresh, dried and processed sweetpotato products, *Ecol. Food Nutr.* **37**(5), 455--473, 1998.
416. Harborne, J.B., & Williams, C.A. Advances in flavonoids research since 1992, *Phytochemistry* **55**, 481--504, 2000.
417. Chen, W., et al. Genome-wide association analyses provide genetic and biochemical insights into natural variation in rice metabolism, *Nat. Genet.* **46**, 714--721, 2014.
418. Attrapadung, S., et al. Identification of ricinoleic acid as an inhibitor of Ca^{2+} signal-mediated cell-cycle regulation in budding yeast, *FEMS Yeast Res.* **10**, 38--43, 2010.
419. Wright, A.J. & Marangoni, A.G. Vegetable oil-based ricinelaidic acid organogels—phase behavior, microstructure and rheology, in *Edible oleogels: structure and health implications*, A.G. Marangoni et al, eds., AOCS Press, Urbana, 2011, 81-99.
420. Walaszek, Z. Potential use of D-glucaric acid derivatives in cancer prevention, *Cancer Lett.* **54** (1-2), 1--8, 1990.
421. Graf, E. Antioxidant potential of ferulic acid, *Free Radic. Biol. Med.* **13**, 435--448, 1992.
422. Lin, F.H., et al. Ferulic acid stabilizes a solution of vitamins C and E and doubles its photoprotection of skin, *J. Invest. Dermatol.* **125**, 826--832, 2005.
423. Iraj, F., et al. Efficacy of topical azelaic acid gel in the treatment of mild-moderate acne vulgaris, *Indian J. Dermatol. Venereol. Leprol.* **73**, 94--96, 2007.
424. Juan, M.E., et al. Antiproliferative and apoptosis-inducing effects of maslinic and oleanolic acids, two pentacyclic triterpenes from olives, on HT-29 colon cancer cells, *Br. J. Nutr.* **100**, 36--43, 2008.
425. Brufau, G., et al. Phytosterols: physiologic and metabolic aspects related to cholesterol-lowering properties, *Nutr. Res.* **28**, 217--225, 2008.

426. Woyengo, T.A., et al. Anticancer effects of phytosterols, *Eur. J. Clin. Nutr.* **63** (7), 813--820, 2009.
427. Sujatha, S., et al. Biological evaluation of (3b)-STIGMAST-5-EN-3-OL as potent anti-diabetic agent in regulating glucose transport using *in vitro* model, *Int. J. Diabetes Mellit.* **2**, 101--109, 2010.
428. Panda, S., et al. Thyroid inhibitory, antiperoxidative and hypoglycemic effects of stigmasterol isolated from *Butea monosperma*, *Fitoterapia* **80** (2), 123--126, 2009.
429. Szuster-Ciesielska, A., et al. Betulin, betulinic acid and butein are inhibitors of acetaldehyde-induced activation of liver stellate cells, *Pharmacol. Rep.* **63**, 1109--1123, 2011.
430. Zhao, C., et al. Extraction of solanesol from tobacco (*Nicotiana tabacum* L.) leaves by bubble column, *Chem. Eng. Process.* **48**, 203--208, 2009.
431. Melo, C.M., et al. α , β -amyrin, a natural triterpenoid ameliorates L-arginine-induced acute pancreatitis in rats, *World J. Gastroenterol.* **16** (34), 4272, 2010.
432. Buckheit, R.W. Jr., et al. The structure-activity relationships of 2,4 (1H,3H)-pyrimidinedione derivatives as potent HIV type 1 and type 2 inhibitors, *Antivir. Chem. Chemother.* **18**, 259--275, 2007.
433. Carlezon, Jr. W.A., et al. Antidepressant-like effects of uridine and omega-3 fatty acids are potentiated by combined treatment in rats, *Biol. Psychiatry.* **57**, 343--350, 2005.
434. Chang, C. I., et al. Arginase modulates nitric oxide production in activated macrophages, *Am. J. Physiol.* **274**, H342--348, 1998.
435. Morris Jr, S. M. Recent advances in arginine metabolism: roles and regulation of the arginases, *Br. J. Pharmacol.* **157**, 922--930, 2009.
436. Soini, J., et al. Norvaline is accumulated after a down-shift of oxygen in *Escherichia coli* W3110, *Microb. Cell Fact.* **7**, 30, 2008.
437. AL-Homeidan, H. H. Application of L-Norvaline for controlling botrytis cinerea on lettuce plant, *Adv. Biol. Res.* **1** (5--6), 159--163, 2007.
438. Smith, E., et al. Isopimaric acid from *Pinus nigra* shows activity against multidrug-resistant and EMRSA strains of *Staphylococcus aureus*, *Phytother. Res.* **19**, 538--542, 2005.

439. Leung, K. N., et al. Immunomodulatory effects of esculetin (6, 7-dihydroxycoumarin) on murine lymphocytes and peritoneal macrophages, *Cell. Mol. Immunol.* **2**, 181--188, 2005.
440. Chin, Y.P., et al. Synthesis and Evaluation of Antibacterial Activities of 5, 7-Dihydroxycoumarin Derivatives, *Arch. Pharm.* **344**, 386--393, 2011.
441. Sapana, S. K., et al. Development and Validation of HPTLC Method for Determination of 3-Hydroxy Androstane [16, 17-C] (6'-methyl, 2'-1-hydroxy-isopropene-1-yl) 4, 5, 6 H Pyran in Jambul Seed (*Syzygium cumini*), *International Journal of PharmTech Research* **1**, 1129--1135, 2009.
442. Kadegowda, A.K.G., et al. *Cis-9, trans-11* Conjugated linoleic acid is endogenously synthesized from palmitelaidic (C16:1 *trans-9*) acid in bovine adipocytes, *J. Anim. Sci.* **91** (4), 1614--1623, 2013.
443. Kelly, G.S. Squalene and its potential clinical uses, *Altern. Med. Rev.* **4**, 29--36, 1999.
444. Weckwerth, W., & Fiehn, O. Can we discover novel pathways using metabolomic analysis? *Curr. Opin. Biotechnol.* **13**, 156--160, 2002.
445. Fiehn, O. Combining genomics, metabolome analysis, and biochemical modeling to understand metabolic networks, *Comp. Funct. Genomics* **2**, 155--168, 2001.
446. Padda, M.S., & Picha, D.H. Antioxidant activity and phenolic composition in 'Beauregard' sweetpotato are affected by root size and leaf age, *J. Amer. Soc. Hortic. Sci.* **132**, 447--451, 2007.
447. Dao, T.T.H. *Metabolic changes in Arabidopsis thaliana plants overexpressing chalcone synthase*. Ph. D. Thesis, Leiden University at Leiden, Netherlands, 1976.
448. Bocobza, S., et al. Riboswitch-dependent gene regulation and its evolution in the plant kingdom, *Genes Dev.* **21**, 2874--2879, 2007.
449. Chen, H., et al. Genetic analysis of pathway regulation for enhancing branched-chain amino acid biosynthesis in plants, *Plant J.* **63**, 573--583, 2010b.
450. Datta, P., & Gest. H. Control of enzyme activity by concerted feedback inhibition, *Proc. Nat. Acad. Sci. U.S.A.* **52**, 1004--1009, 1964.
451. Carvalho, I. S., et al. Effect of photoperiod on flavonoid pathway activity in sweet potato (*Ipomoea batatas* (L.) Lam.) leaves, *Food Chem.* **118**, 384--390, 2010.

452. Wang, H., et al. Functional characterization of Dihydroflavonol-4-reductase in anthocyanin biosynthesis of purple sweet potato underlies the direct evidence of anthocyanins function against abiotic stresses, *PLoS ONE* **8**, e78484, 2013.
453. Gayen, D., et al. Comparative analysis of nutritional compositions of transgenic high iron rice with its non-transgenic counterpart, *Food Chem.* **138**, 835--840, 2013.
454. Harrigan, G.G., et al. Impact of genetics and environment on nutritional and metabolite components of maize grain, *J. Agric. Food Chem.* **55**, 6177--6185, 2007.
455. Jones, D.H. Phenylalanine ammonia-lyase: regulation of its induction, and its role in plant development, *Phytochemistry* **23**, 1349--1359, 1984.
456. Ruiz-García, Y., & Gómez-Plaza, E. Elicitors: A tool for improving fruit phenolic content, *Agriculture* **3**, 33--52, 2013.
457. Wink, M. Introduction: Biochemistry, physiology and ecological functions of secondary metabolites, in *Annual Plant Reviews Volume 40: Biochemistry of Plant Secondary Metabolism*, 2nd ed., M. Wink, eds., Wiley-Blackwell Publishing, New York, 2010. 1--17.
458. Taiz, L. & Zeiger, E. Stress Physiology, in *Plant Physiology*, R.A. Bressan et al, eds., Sinauer Associates Press, Sunderland, 1998, 591--614.
459. Ghasemzadeh, A., & Ghasemzadeh, N. Flavonoids and phenolic acids: Role and biochemical activity in plants and human, *J. Med. Plant Res.* **5** (31), 6697--6703, 2011.
460. Cartea, M.E., et al. Phenolic Compounds in *Brassica* Vegetables, *Molecules* **16** (1), 251--280, 2011.
461. Perales-Sánchez, J.X.K., et al. Increasing the antioxidant activity, total phenolic and flavonoid contents by optimizing the germination conditions of amaranth seeds, *Plant Foods Hum. Nutr.* **69**, 196--202, 2014.

Bacterial strains and Plasmids

Strain	Genotype
<i>Escherichia coli</i> DH5 α	Φ 8dlacZ Δ M15, recA1, endA1, gyr A96, thi-1, hsd17 (r_k^- , m_k^-) supE44, relA1, deoR, (LacZYA-argF)U19
Helper strain -HB101:: pRK 2013 (Clontech)	
<i>Agrobacterium tumefaciens</i> (EHA105)	A virulent strain carrying pMP90 Ti-plasmid conferring resistance against gentamicin and rifampicin as chromosomal selection
Plasmid	pBI121 (Clontech), pSB 8, pSB8 β

Media and Solutions

YEP:	1% Yeast Extract 1% Bacto Peptone 0.5% NaCl
Luria Broth (LB):	25 g/l
LB agar:	32 g/l
Micropropagation medium (MM):	MS medium 0.1 mg/l IAA, pH 5.6-5.8
Callus induction medium (CIM):	Gamborg's B-5 medium 0.4 mg/l NAA, pH 5.6-5.8
Shoot induction medium (SIM):	Gamborg's B-5 medium 0.1-0.4 mg/l NAA, pH 5.6-5.8
Linsmaier and Skoog medium:	LS medium
TE (pH 8.0):	10 mM Tris-HCl (pH 8.0) 1 mM EDTA (pH 8.0)
20X SSC:	175.3 g NaCl 88.2 g Trisodium citrate pH 7.0

6X Endo-R:	30% Ficoll 400 60 mM EDTA (pH 8.0) 0.6% SDS
High Salt TE:	10 mM Tris (pH 8.0) 1 mM EDTA
Fixative:	0.3% paraformaldehyde 10 mM MES pH5.6 0.3 M Mannitol 50 mM NaH ₂ PO ₄ (pH 7.0)
20X SSC (1 liter):	175.3 g Sodium Chloride 88.2g Sodium Citrate, pH 7.0
10X MOPS Buffer:	200 mM 3-[<i>N</i> -morpholino] propanesulfonic acid (MOPS) 50 mM sodium acetate 10 mM EDTA final pH of 6.5–7.0 with NaOH
RNA loading dye:	95% formamide 0.025% SDS 0.025% bromophenol blue 0.025% xylene cyanol FF 0.025% ethidium bromide
2X Extraction Buffer:	50 mM Tris (pH 6.8) 1% 2-mercaptoethanol 1 mM PMSF 1 mM EDTA
4X Stacking Buffer:	0.5 M Tris HCl (pH 6.8)
8X Resolving Buffer:	3 M Tris HCl (pH 8.8)

12.5% polyacrylamide gel:	Acrylamide (30:0.8) - 16.68 ml
(40 ml)	4X buffer (pH 8.8) - 10 ml
	MilliQ - 12.92 ml
	SDS (20%) - 200 µl
	APS (10%) - 200 µl
	TEMED - 15 µl
Reservoir Buffer (10X):	0.25 M Tris (pH 8.3)
	1.92 M Glycine
	1% SDS
Towbin's Buffer:	25 mM Tris
	190 mM Glycine
	20% Methanol
TBS:	10 mM Tris-HCl, pH 8.0
	100 mM Tris, pH 8.0
	150 mM NaCl
TBST:	TBS, 0.05% Tween 20
Blocking Solution:	TBS + 5% Fat free dry milk
AP Buffer:	100 mM Tris-Cl, pH 9.5
	100 mM NaCl
	50 mM MgCl ₂
AP Colour Development Solution:	10 ml AP Buffer
	66 µl NBT (50 mg/ml, 70% DMF)
	33 µl BCIP (50 mg/ml DMF)
X-gal:	20 mg/ml (in DMF)
IPTG:	200 mg/ml in H ₂ O
Antibiotics:	Kanamycin 100 mg/ml of water
	Rifampicin 50 mg/ml of methanol
	Cefotaxime 250 mg/ml of water
	Paromomycin sulfate 50mg/l of water

List of primers:

Genes	Primer sequence
<i>nptIIF</i>	5' - ATGATTGAACAAGATGGATTGCACGCAGG -3'
<i>nptIIR</i>	5' - GAAGAACTCGTCAAGAAGGCGATA -3'
<i>AmAIF</i>	5'-CACCATGGCGGGATTACCAGTG-3'
<i>AmAIR</i>	5'-CAAGGAAGAACCCTCTTGTTTCC-3'
<i>TubRTF</i>	5' - AGGACCCTTGTGTTTGGTGTTAA- 3'
<i>TubRTR</i>	5' - CCCACTCATCGTTGCAGAAA-3'
<i>GAPDHRTF</i>	5' - AAGAAAACAAAAGCACGGCACTA-3'
<i>GAPDHRTR</i>	5' - AAGTGGAAAAAGGATTCGGTGTAT-3'
<i>ActinF</i>	5'-CTCCCCTAATGAGTGTGATGTGAT-3'
<i>ActinR</i>	5' - GAGCCCCATGAGAACATTACCA-3'
<i>GUSF</i>	5'-TGGTAATTACCGACGAAAACGGC-3'
<i>GUSR</i>	5'-ACGCGTGGTTACAGTCTTGCG-3'
<i>AmAIRTF</i>	5'-GGGAATGATCCTCGCGAAA-3'
<i>AmAIRTR</i>	5' - AAAATCATGCACATCCGACCTA-3'
<i>AmAIUTRRTF</i>	5' - GAGATAATAGAATTGGGATCCAACAAC-3'
<i>AmAIUTRRTR</i>	5' - CCAAAGAGACGACTTACAACGTTTT-3'

List of chemicals:

Type	Material	Source
Antibiotics	Ampicillin, Kanamycin, Rifampicin, Spectinomycin, Geneticin	Sigma
Disposable filters	PVDF 0.45 µm filter unit	Millipore
Enzymes	Commonly used restriction enzymes	NEB
	<i>Taq</i> DNA Polymerase	Clontech, Finnezym
	T4 DNA Ligase	NEB
	RNase	BioBasic, Amersham
Dyes	Ethidium Bromide, Xylene cyanol Methylene Blue, Coomassie Brilliant Blue	Amersham
Culture media components	Tryptone, Yeast Extract, Agar, MS salts	Difco, Invitrogen, Sigma
Locally available chemicals	Isopropanol, iso-amyl alcohol, CaCl ₂ , NaCl, NaOH, Glucose, Methanol, MgCl ₂ , KOH, Potassium acetate, Chloroform, Glycerol, Acetic acid, NaH ₂ PO ₄ , Na ₂ HPO ₄ , MgSO ₄ , HCl, H ₂ SO ₄ , Glycine, KCl, Sucrose, Pot. Dichromate, Sodium hypochlorite, Mercuric chloride, tri-Sodium citrate, Formaldehyde.	Qualigens and Merck
Foreign chemicals	DEPC, HEPES, IPTG, MOPS, Sephadex G-50, EDTA, CTAB, Acrylamide, Bis-Acrylamide, TEMED, Spermine, Spermidine, Polyvinyl Polypyrrolidone, Triton-X-100, X-gal	Amersham, Sigma, Ambion,