# **Chapter-1** Introduction

# 1. Introduction

A multitude of parasites, spanning from protozoa to helminths, have the potential to endanger human health. Parasites responsible for various diseases are characterized by heightened levels of both morbidity and mortality. To show its influence, diverse parasitic species employ a range of infection mechanisms across various host species [1]. This variation contributes to the adaptability and complexity of parasitic infections in different hosts. Certain parasitic organisms which cause diseases like Chagas, Leishmaniasis, Schistosomiasis, etc., fall under the classification of infectious diseases known as Neglected Tropical Diseases (NTDs). The data indicates that NTDs are primarily prevalent in countries across Africa, Asia, and some parts of South America [2].

## 1.1. Leishmaniasis- An overview

Leishmaniasis is one of the NTDs which affects many countries across the globe and it stands out as a crucial parasitic ailment, contributing substantially to global mortality and morbidity. An intracellular protozoan parasite belonging to the *Leishmania* genus is required to spread this disease through a vector [3]. Leishmaniasis are believed to impact approximately 0.7–1 million individuals, with a global population of 350 million people facing the risk of infection. According to global data provided by the World Health Organization (WHO), there are over 12 million individuals currently infected, with an annual incidence of new cases ranging between 0.9 to 1.6 million. The disease leads to an estimated 20,000 to 30,000 deaths annually, and a staggering 350 million people are at risk of contracting the infection [4]. Moreover, among the 30 identified species of the *Leishmania* parasite to date, 21 have been documented as responsible for causing the disease known as leishmaniasis in humans. These 21 species of *Leishmania* contribute to distinct forms of leishmaniasis, namely cutaneous, mucocutaneous, and visceral leishmaniasis [3].

# 1.2. Etiology

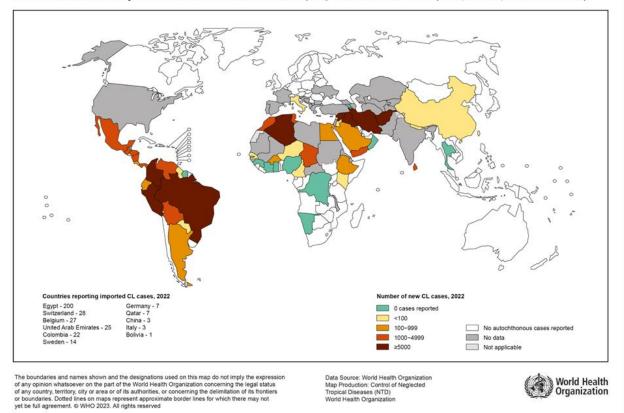
Leishmaniasis results from the infection caused by the *Leishmania* parasite, with the female sand-fly Phlebotomus acts as the vector [5]. The initiation of leishmaniasis disease occurs when parasites are conveyed to the mammalian host by the means of blood-feeding activity of an infected female sandfly. The parasites, existing in two forms, enter the human host as metacyclic promastigotes, and subsequently undergo a transformation into their immobile amastigote form [6]. The array of *Leishmania* species as mentioned, includes *L. donovani, L. major, L. braziliensis, L. infantum, L. mexicana,* and *L. chagasi* that gives rise to various

forms of Leishmaniasis. The spectrum of disease severity encompasses individuals who are serologically positive but asymptomatic, all these cases marked by disfiguration and the potential for fatal outcomes [7]. Based on the clinical presentation, leishmaniasis is categorized into four primary forms. The most life-threatening form of leishmaniasis, Visceral Leishmaniasis (VL), is caused by L. donovani and L. infantum. When VL establishes itself in the host, symptoms include the enlargement of the liver and spleen, accompanied by anemia, weight loss, and intermittent fevers [8]. In context of India, individuals affected by VL exhibit skin darkening, a phenomenon attributed to the cytokinetriggered synthesis of adrenocorticotrophic hormone. Thus, VL is considered as "kala-azar" in India [9]. Cutaneous leishmaniasis (CL), a non-fatal variant of leishmaniasis, is prominently caused by numerous Leishmania species, including but not limited to L. major, L. tropica, L. amazonesis, L. braziliensis, L. mexicana, L. panamensis, and others. The lesions occurred by CL are confined to the site and may undergo self-healing [10]. Nevertheless, the visual characteristics and duration of healing differ among species, often resulting in notable scarring and an extended healing period spanning several years. Mucocutaneous leishmaniasis (MCL), a less common form of leishmaniasis, manifests on the mucous membranes of the face, including the nose, mouth, and throat. In this form, parasites can access the mucosa either through the dissemination via the lymphatic system or by direct injection into the region through the bite of an infected sandfly [11]. Manifestations of this leishmaniasis encompass symptoms such as nasal discharge, congestion, and episodes of sudden nosebleeds, which may manifest years following a starting cutaneous lesion [12].

#### 1.3. Epidemiology

The prevalence of leishmaniasis exhibits a broad geographical distribution, with reported cases spanning the globe. Regions characterized by inadequate sanitation and substandard living conditions foster an elevated proliferation of vectors, contributing to a higher incidence of cases, particularly in developing nations [13, 14]. As of November 2023, the WHO released a report detailing the global endemic status of CL of 2022. Currently, approximately 95% of endemic cases of the cutaneous form are concentrated in regions including America, the Mediterranean basin, the Middle East, and countries in central Asia. Some of the countries where the cutaneous form is endemic are Brazil, Colombia, Ethiopia, Sudan, Afghanistan, Iran, Pakistan, Tunisia, etc., [15]. Regarding VL, WHO presented the worldwide endemic situation in 2022. The report highlighted countries like India, South

Sudan, Sudan, Brazil, Ethiopia and Somalia as having the highest incidence of VL cases [15]. The prevalence of VL is disproportionately concentrated in these nations, accounting for 90% of all reported cases. VL is extensively documented in numerous countries, with India and Sudan being a significant contributor to the global burden [16]. India accounts for 18% of the global burden of VL in 2020, emphasizing its substantial impact on the prevalence of the disease [17]. In India, the primary states reporting VL cases are Bihar, Jharkhand, West Bengal and Uttar Pradesh, according to information obtained from the National Vector Borne Disease Control Programme (NVBDCP) of India [18]. MCL has been documented in regions spanning South America, Asia, Europe and Africa. Noteworthy countries reporting MCL cases include India, Sri Lanka, Pakistan, Iran, Saudi Arabia and Sudan [19]. WHO presented the global endemic situation of CL and VL in 2022 through Figures 1.1 and 1.2. Additionally, Figure 1.3 depicted the distribution of different species of Leishmaniasis in Indian subcontinent.



Status of endemicity of cutaneous leishmaniasis (CL) worldwide, 2022 (as reported by November 2023)

#### Figure 1.1: Endemic status of CL 2022 taken from WHO

Status of endemicity of visceral leishmaniasis (VL) worldwide, 2022 (as reported by November 2023)

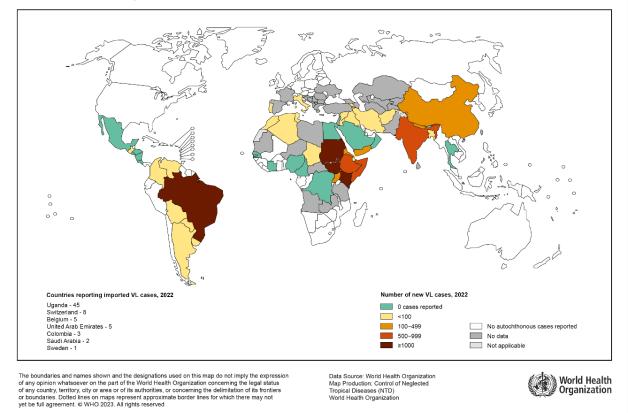


Figure 1.2: Endemic status of VL 2022 taken from WHO

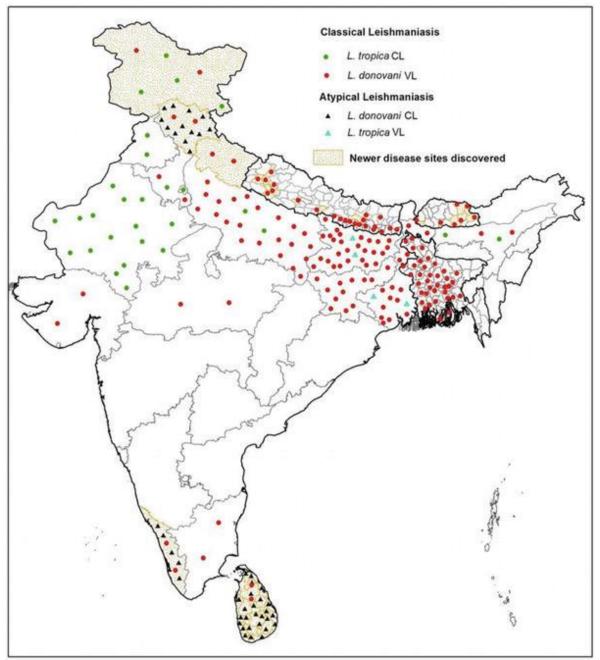


Figure 1.3: Distribution of Leishmaniasis in Indian subcontinent [17].

# 1.4. Leishmaniasis treatment

Like many other diseases, leishmaniasis possesses treatment options, yet these therapeutics encounter multifaceted challenges, including elevated costs, efficacy concerns, toxicity and potential side effects [20]. Moreover, the rise of drug resistance, coupled with a limited understanding of the interplay between the host and the pathogen, complicates the efficient implementation of treatment strategies. The limited knowledge of immunobiology of the parasite presents a substantial hurdle in the quest for new drugs and vaccines for this disease. Currently, various treatment options for leishmaniasis remain under scrutiny, awaiting further exploration and insights [21].

#### 1.4.1. Chemotherapy

Chemotherapy is a treatment method for leishmaniasis that utilizes chemical substances, usually in the form of drugs or medications, to treat a disease. These drugs are employed to combat the parasitic infection caused by Leishmania species, aiming to eliminate or suppress the growth of the parasites within the host's body [22]. The therapeutic arsenal for leishmaniasis encompasses medications such as pentavalent antimony, amphotericin B (AmpB), miltefosine, paromomycin, and pentamidine [23]. While pentavalent antimony stands as the primary treatment for VL and CL, its notable drawback lies in the occurrence of side effects upon administration. These side effects, including vomiting, nausea, headache, and anorexia, present a challenge in the pursuit of effective and well-tolerated treatments for the disease [24]. In addition to its side effects, resistance to pentavalent antimony has emerged, resulting in therapeutic challenges and treatment failures. Factors contributing to this resistance include a compromised host defense system, the use of unauthorized drug batches, the specific *Leishmania* species involved, and incomplete drug treatment durations [25]. AmpB has risen as a secondary line of treatment. However, the widespread clinical application of AmpB has been restricted owing to its considerable toxicity. This high toxicity level can induce clinical complications such as nephrotoxicity, liver damage, nausea, fever, etc., in patients [26]. The drug's mechanism of action occurs within the cytoplasmic membrane of parasites, where it specifically binds to ergosterol. This interaction facilitates an increase in membrane permeability and promotes ion influx [27]. AmpB stands as the most efficacious treatment presently available for VL in India. Additionally, it serves as a treatment option for individuals with co-infections of VL along with HIV or tuberculosis (TB) [28]. Miltefosine stands as the solitary oral medication effective in treating both VL and CL, with a prescribed treatment duration of 28 days. However, a significant drawback is its prolonged elimination process, resulting in an extended residence time in the body. This extended duration elevates the risk of side effects for patients undergoing the drug regimen [29]. Combined therapeutic approaches involving liposomal AmpB with miltefosine, as well as miltefosine with paromomycin, have been employed for treating VL. This combination strategy has demonstrated enhanced costeffectiveness compared to the use of miltefosine alone [30]. Paromomycin, an aminoglycoside antibiotic, demonstrates heightened efficacy against CL and is also

employed in the treatment of VL. Despite the current lack of clarity on the precise mechanism underlying paromomycin's action in the *Leishmania* pathogen, its notable capacity for inhibiting various *Leishmania* species is evident [31]. Pentamidine serves as a treatment for VL when resistance develops to primary or secondary therapeutic options. Administered via the intramuscular route, the exact mechanism of action of pentamidine is unclear, but it is known to affect the parasitic mitochondria. However, instances of resistance to pentamidine have been reported in certain cases of leishmaniasis [32]. Table 1.1 provides an overview of various chemotherapeutic drugs, detailing their routes of administration along with their respective advantages and disadvantages.

Sl. No.	Drug	Route of administration	Pros	Cons
1	Pentavalent antimonial	Intralesional, intramuscular or intravenous	Cheap and easily available	Develop resistance
2	AmpB	Intravenous	Less resistance	Nephrotoxicity
3	Liposomal AmpB	Intravenous	Less toxic	Costly drugs
4	Miltefosine	Oral	Better efficacy	Drug resistance
5	Paromomycin	Intramuscular (VL) or topical (CL)	Low cost	Efficacy varies with species and region
6	Pentamidine	Intramuscular	Small course of treatment	Efficacy of drug changes with species

## Table 1.1: Chemotherapeutic drugs for leishmaniasis treatment [20].

#### 1.4.2. Immunotherapy

Immunotherapy for leishmaniasis entails leveraging the body's immune system to combat the parasitic infection induced by *Leishmania* species. The objective of this therapeutic approach is to augment or adjust the immune response, facilitating improved control and elimination of the parasites [33]. The primary idea of this therapy is to diminish the adverse effects of the drug on the host and to mitigate the development of resistance. Diverse immunotherapeutic strategies were involved which includes the utilization of cytokines, immunomodulators, the amalgamation of chemotherapy and immunotherapy, as well as vaccines, have been implemented in the management of both CL and VL [34]. Immunotherapy centered around cytokines has attracted considerable attention due to its potential role in preventing resistance. Key cytokines in this context include Interleukin (IL)-12 and interferon (IFN)- $\gamma$ , which play pivotal roles in a variety of immunotherapeutic strategies. The intricate balance between pro- and anti-inflammatory cytokines holds paramount importance in the immunopathogenesis of Leishmania infection [35]. Immunomodulators in the context of leishmaniasis denote compounds or agents capable of modulating and regulating the immune response in the presence of Leishmania parasites. The impact of these immunomodulators on both VL and CL has been investigated, either as standalone interventions or in conjunction with chemotherapeutic agents. This exploration aims to discern the nuanced effects of immunomodulation on the immune system's response to *Leishmania* infection, contributing to a more comprehensive understanding of potential therapeutic strategies [36]. The combined application of chemotherapy and immunotherapy for CL and VL demonstrated superior efficacy compared to individual therapeutic modalities. In murine models, the implementation of a combination therapy approach resulted in a notable reduction in CL lesions. Similarly, for VL, the combination therapy exhibited an augmented rate of recovery [33]. Numerous studies have delved into varied components integrated into vaccine formulations, encompassing specific elements derived from Leishmania, to serve as immunotherapeutic agents. Moreover, distinct constituents, such as specific leishmanial components, live parasites, and inactivated parasites, has been explored in diverse research endeavors as immunotherapeutic tools. Several potential vaccine candidates underwent testing in experimental models, including dogs, BALB/c mice, and hamsters, to evaluate their efficacy against *Leishmania* infection [35]. The elicitation of immunity following infection implies that vaccination holds a viable strategy for preventing cutaneous leishmaniasis, both in animal models and human subjects [37].

#### 1.4.3. Nanoparticles therapy

In the medical domain, nanoparticles have emerged as pivotal therapeutic entities. Their application in pathogen elimination involves leveraging inhibitory effects. Nanoparticles, whether utilized independently or in conjunction with other agents, demonstrate the potential to eliminate parasites. Studies have indicated that the co-administration of nanoparticles with drugs enhances efficacy while concurrently mitigating adverse effects [38]. Nanoparticles generally provide various benefits, including reduced toxicity, enhanced

delivery systems, improved solubility for hydrophobic activation, and increased bioavailability. In the treatment of leishmaniasis, different structural nanocomponents were utilized which encompass liposome nanoparticles, polymeric nanoparticles, lipid nanoparticles, as well as TiO<sub>2</sub>, Ag<sub>2</sub>O, and ZnO nanoparticles [39]. Exploration of nanoparticle-based drug studies on *Leishmania* species has expanded and improved the treatment options for leishmaniasis, paving the way for more effective anti-leishmania drugs.

# i. Liposome nanoparticles

Liposomal nanoparticle systems exhibit regulated drug release, imparting heightened treatment efficacy. The adsorption or encapsulation of drugs within nanoparticles showed antileishmanial activity and results in a significant reduction in toxicity, minimizing adverse effects on the host. Additionally, this mechanism facilitates enhanced drug delivery in comparison to conventional formulations in leishmaniasis treatment [40]. Medications employing liposome nanostructures for the treatment of different manifestations of leishmaniasis include liposomes containing hexadecylphosphocholine, liposomal AmpB, bovine serum albumin nanoparticles incorporating AmpB, mannosylated chitosan nanoparticles loaded with curcumin, chitosan nanoparticles conjugated with mannose and lipid formulations of AmpB [39].

# ii. Polymeric nanoparticles

Biodegradable polymer nanoparticles find extensive utilization in drug delivery and are also employed for encapsulating essential molecules such as DNA, RNA, and proteins, playing a pivotal role in biomedical applications [41]. Various potential drugs related to leishmaniasis use polymer nanoparticles for showing anti-leishmanial activities. Some of the polymeric based nanoparticles are Primaquine-loaded polyisohexylcyanoacrylate nanoparticles [42], AmpB-loaded nanoparticles [43], mannose-anchored thiolated AmpB nanocarriers [44], mannosylated thiolated paromomycin-loaded PLGA nanoparticles [45], etc.

# iii. Lipid nanoparticles

Solid lipid nanoparticles (SLNs) are composed of biocompatible lipids that maintain a solid state at room temperature. These carriers offer benefits such as controlled drug release, improved stability, and the capacity to encapsulate pharmaceutical agents. Conversely,

nanostructured lipid carriers (NLCs) involve a combination of solid and liquid lipids, forming a more flexible matrix with additional features like enhanced drug loading, controlled release, and increased stability compared to SLNs [46]. Both SLNs and NLCs are recognized as second-generation colloid carriers, primarily employed to augment drug delivery. Notably, these nanoparticle carriers have been employed in anti-parasitic drugs to enhance efficacy [47]. The application of conjugated AmpB with SLNs and paromomycin with SLN has been investigated in leishmanial infections, revealing heightened effectiveness [48, 49]. Lipid nanoparticles containing oryzalin, artemisinin-loaded poly lactic co-glycolic acid nanoparticles, biogenic silver nanoparticles, and andrographolide nanoparticles are among the lipid-based nanoparticles utilized in various model studies infected with *Leishmania* species, demonstrating superior anti-leishmanial effectiveness [39].

## iv. TiO2 and Ag2O nanoparticles

Distinctive physical and chemical properties of titanium dioxide (TiO<sub>2</sub>) and silver oxide (Ag<sub>2</sub>O) nanoparticles have captivated numerous researchers, prompting their utilization in medicine and therapeutic applications. These two nanoparticles of TiO<sub>2</sub> and Ag<sub>2</sub>O was reported to show antimicrobial solution on bacteria and *Leishmania* parasite, thus showed antileishmanial effect [50]. A study that combined application of meglumine antimoniate with TiO<sub>2</sub>Ag nanoparticles intensifies its anti-leishmanial efficacy, concurrently mitigating the drug's toxicity [51]. In a research investigation, the proliferation and metabolic activity of *Leishmania* parasites are suppressed by silver nanoparticles. When exposed to UV light, Ag<sub>2</sub>O nanoparticles exhibit superior augmented antimicrobial efficacy [52].

#### 1.5. Development of vaccines against Leishmaniasis

Currently, there are no licensed vaccines available for the prevention of leishmaniasis, despite the significant impact of the *Leishmania* parasite on people, including deaths and adverse effects. However, vaccine development efforts have been underway for several decades, aiming to provide long-term protection against this debilitating disease. Leishmaniasis treatment depends on existing anti-leishmanial medications, yet they fall short due to their expense, toxicity, and adverse effects, as well as prolonged dosing regimens with varying levels of effectiveness. Moreover, anti-leishmanial drugs currently face resistance in multiple endemic regions worldwide, exacerbating the situation. Consequently, there is an instant need to create a potential vaccine against leishmaniasis. In this context, various vaccine candidates and strategies have been explored, ranging from live

attenuated and subunit vaccines to DNA and recombinant protein-based approaches. Understanding the challenges and advancements in vaccine development for leishmaniasis is crucial for addressing the unmet medical need posed by this neglected tropical disease. Table 1.2 presents various generations of vaccines, types of vaccines, methods, and expressions used in combating leishmaniasis.

# 1.5.1. First Generation Vaccines of Leishmaniasis

First-generation vaccines for combating leishmaniasis typically leverage antigens sourced from the entirety of the parasite, whether in an inactivated state or attenuated for live use. The ease and cost-efficiency of cultivating Leishmania in cell-free media facilitated the initial adoption of whole inactivated parasites [53]. These pioneering vaccines, originating from early investigations dating back to the late 1930s and 1940s, aimed to instigate protective immune responses against *Leishmania* infections. Notably, a study conducted by Mayrink et al. in Brazil delved into the development and evaluation of a first-generation vaccine against leishmaniasis, yielding satisfactory outcomes albeit with only a 50% protective effect [54]. Further research in Brazil underscored the efficacy of a combined treatment approach involving killed L. amazonensis paired with a partial dose of meglumine antimoniate for addressing CL [55]. Additionally, investigations in Ecuador revealed promising protection against CL through the utilization of a formulation comprising L. brasiliensis, L. guianensis, and L. amazonensis [56]. In Venezuela, a combination of adjuvant Bacillus Calmette-Guérin (BCG) with killed L. amazonensis was employed to treat CL [57]. Furthermore, the adjunctive application of BCG alongside killed Leishmania vaccines was investigated to bolster the cell-mediated response in experimental models [58]. In Iran and Sudan, examinations have assessed a killed vaccine derived from L. major, manufactured by the Razi Institute [58,59]. These inquiries highlight the potential of employing a killed Leishmania vaccine, with or without BCG as an adjuvant, as a safe alternative [60]. The safety aspect of killed vaccines underscores their favorable profile in terms of safety, rendering them a promising avenue for further exploration in vaccine development endeavors.

Live-attenuated vaccines involve intentionally removing genes essential for an organism's virulence and survival. This modification enables them to stimulate an immune response similar to a natural infection. Although live-attenuated vaccines closely mimic natural infection and effectively induce protective immunity, concerns about safety hinder their full

potential benefits. Existence of antibiotic-resistant genes, and the risks involved in vaccinating immunosuppressed individuals are significant factors to address. Vaccines derived from *L. major, L. mexicana, L. braziliensis,* and *L. donovani* have been created, demonstrating notable efficacy in safeguarding susceptible mice against CL and VL [61, 62,63]. A genetically modified strain of *L. donovani* was developed through the deletion of the gene responsible for Ldp27. When immunized with these Ldp27-deficient parasites, subjects exhibited diminished parasitic burdens and experienced improved long-term Th1-mediated protection against challenges of CL and VL [64].

#### 1.5.2. Second generation vaccines of Leishmaniasis

Second-generation vaccines center on pathogen antigens, comprising purified fractions of native proteins, genetically engineered subunits. These vaccines consist of precisely defined molecular constituents, employing recombinant antigens—potentially one or more proteins-and designated adjuvants [65]. In the context of Leishmania, factors such as antigenic variation, antigen conservation among different species, and accessibility of the Leishmania genome sequence are taken into account. This has allowed for the utilization of Leishmania proteins as potential vaccine applicants [66]. The approach relies on utilizing recombinant subunits alongside modified Leishmania strains, recombinant bacteria or viruses harboring genes encoding *Leishmania* antigens [67]. The Infectious Disease Research Institute (IDRI) reached a significant milestone by developing LEISH-F1, a pioneering recombinant vaccine candidate that successfully advanced to clinical trials. Additionally, IDRI has developed candidates like LIESH-F2 and LEISH-F3, both of which have progressed to clinical trials, showcasing encouraging outcomes [68]. Studies have investigated the potential of utilizing sand fly salivary proteins as a standalone vaccine against leishmaniasis, or in combination with Leishmania proteins, to create a potent vaccine candidate. Research suggests that individual saliva antigens may exert paradoxical impacts on disease outcomes when compared to whole saliva. Additionally, findings have revealed that salivary proteins can independently initiate the cell-mediated immune response, unlike recombinant protein candidates that often rely on adjuvants for activation [69].

#### 1.5.3. Third generation vaccines of Leishmaniasis

As third-generation vaccines, genetic immunization strategies have emerged, utilizing the direct administration of nucleic acids, including mRNA, naked plasmid DNA, or

encapsulated within a viral vector. Third-generation vaccines surpass second-generation counterparts by providing improved protein stability, leading to a significantly elevated level of protection. In murine models, an RNA vaccine expressing LEISH-F2 followed by a subunit vaccine demonstrated effectiveness against L. donovani [70]. These vaccines, containing plasmid DNA, prompt the synthesis of endogenous proteins post-injection, thus eliciting targeted immune responses. They facilitate the development of both cellular and humoral immunity and are offered in various formats, such as recombinant proteins, single vaccines, or multigene formulations [71, 72]. In the realm of Leishmania research, a broad spectrum of antigens has been explored, and commonly used DNA vaccines primarily consist of protein components. Currently, pre-clinical trials for the development of leishmaniasis vaccines involved testing on mouse models infected with L. major, L. mexicana, and L. amazonensis. Moreover, in the evaluation of vaccines, strains of VL including L. donovani, L. infantum, and L. chagasi were utilized in various animal models. [65]. The initial DNA vaccine designed for leishmaniasis featured the gene encoding gp63, which has demonstrated its ability to induce a Th1 response and offer substantial protection against CL due to L. major. LACK, another DNA vaccine extensively studied, offers diverse levels of protection against infections caused by L. major, L. donovani, and L. infantum in mouse and dog models [73,74]. DNA vaccines are attractive because of their simple manufacturing process, stability, and various safety characteristics. Moreover, they are acknowledged for their ability to trigger stronger Th1 immune responses in contrast to protein-based vaccines. Despite demonstrating efficacy in animal models, DNA vaccines have faced challenges in clinical trials, particularly in terms of their immunogenicity and clinical benefits. This has made their translation to human use challenging [75, 76]. Currently, there are no approved DNA vaccines for human use, including those targeting leishmaniasis and other infections.

Sl. No.	Generation	Type of vaccine	Methods	Expression
1	First generation vaccine	Killed Leishmania	Formulations of polyvalent	CL
			BCG as adjuvant	CL
		Live attenuated	Biopterin transporter 1 (BT1)-deleted <i>L. donovani</i>	VL
			Ldp27 gene deleted L. donovani	VL
			(HSP)-70-II null L. infantum	VL and CL
			L. infantum KHARON1 (KH1) null	VL
2	Second generation vaccine	Subunit and recombinant vaccines	LEISH-F1, LIESH- F2 and LEISH-F3	VL and CL
			LJM19 and LJL143	VL and CL
			PdSP15	VL and CL
3	Third generation vaccine	DNA vaccines	LEISH-F2	VL
			gp63	VL and CL
			LACK	VL and CL

# Table 1.2: Vaccination strategies investigated for combatting Leishmania [77].

#### 1.6. Drug resistance in Leishmaniasis

Leishmaniasis, a neglected tropical disease endemic in over 98 countries. Treatment failure in patients with Leishmaniasis is a frequent occurrence, as evidenced by cases where the same species of *Leishmania* causing identical clinical manifestations exhibit varying responses to the same drugs, ultimately resulting in treatment inefficacy for affected individuals [78]. Treatment failure in Leishmaniasis can stem from a myriad of reasons, ranging from issues with the drugs themselves to complications like co-infections such as HIV or the presence of *Leishmania* RNA virus. Parasite-related factors encompass various elements, such as the inherent virulence of the specific strain of *Leishmania* causing the infection. Furthermore, leishmaniasis caused by RNA viruses can incite distinct host immune responses. Moreover, treatment failure can occur when individuals with leishmaniasis exhibit favorable responses to a specific therapy for similar clinical manifestations, but this efficacy is not observed universally among all patients. This scenario arises when patients are infected with different *Leishmania* species or strains [79]. Additionally, factors related to the host, like compromised immunity or incorrect dosages, can also contribute [80]. Moreover, the emergence of drug resistance poses a significant challenge in effectively treating this disease.

Drug resistance in Leishmaniasis emerges as the *Leishmania* parasites evolve mechanisms to resist the impact of anti-leishmanial medications, undermining the effectiveness of treatment plans. Across different regions and in the treatment of VL, a variety of antileishmanial drugs are employed. Notably, pentavalent antimonial compounds have historically been administered to treat VL in areas such as Africa, South America, India and Nepal. Conversely, in the Mediterranean basin, liposomal AmpB has been the preferred choice [81]. From many decades, pentavalent antimonials, including sodium stibogluconate and meglumine antimonate, continue to be the primary choice for treating VL. This preference stems from studies demonstrating the heightened sensitivity of *L. donovani* and *L. brasiliensis* to sodium stibogluconate compared to other *Leishmania* species, coupled with their extensive use over two decades [82]. However, the emergence of resistance to pentavalent antimonials was first identified in North Bihar in 1980s, where approximately a quarter of individuals exhibited unresponsiveness to these medications [83].

Both antimony and arsenic are heavy metals, sharing certain characteristics. In the northeastern regions of India, it has been noted that *Leishmania* parasites exhibit reduced responsiveness to antimonial drugs. This phenomenon is attributed to the high levels of arsenic present in the groundwater of this area. Consequently, arsenic exposure to the *Leishmania* pathogen has led to the growth of antimony resistance in certain regions of India. Additionally, individuals afflicted with leishmaniasis residing in areas with lower arsenic levels demonstrate a more favorable response to antimonial drugs [84]. Research has revealed that resistant parasites exhibit lower levels of antimony compared to their sensitive counterparts. Increased expression of the Aquaporin 1 membrane carrier (AQP1) enhances parasite sensitivity to antimonial drugs, whereas decreased levels of AQP1 lead to reduced responsiveness and eventual resistance. Furthermore, deletion or mutation of the AQP1 gene confers resistance to antimonial upon parasite exposure [85,86]. For pentavalent antimony to exert its action within the parasite, it must undergo reduction to its trivalent form. Diminished biological reduction of antimony results in reduced drug uptake, thereby

promoting resistance. Additionally, overexpression of ATP-binding cassette (ABC) transporters such as ABCI4 and ABCG2 facilitates drug efflux, contributing to antimonial resistance [87,88].

Miltefosine (MIL), introduced in India in 2002, emerged as a promising alternative to pentavalent antimonials due to its high cure rates against leishmaniasis [89]. Miltefosine interferes with phospholipid synthesis and metabolism. Furthermore, it facilitates parasite cell death while having a minimal impact on the host, thus enhancing its therapeutic efficacy [90]. Following a decade since its introduction, the efficacy of MIL has shown a notable decline, transitioning from approximately 90% efficacy during its initial implementation to a range of 10-20% in parasites, consequently resulting in treatment failures. Observations indicate that MIL necessitates approximately 120 hours to eliminate half of the administered doses, which is a considerable high duration [91]. Consequently, its prolonged presence in the body exposes individuals to potential side effects and increases the likelihood of resistance development. The Leishmania miltefosine transporter (LMT and/or LRos3) serves as the primary mechanism for the uptake of MIL within the parasite. Experimental evidence from both in vitro and in vivo studies has demonstrated that mutations or deletions occurring in the LMT gene result in an increase in parasite resistance [92]. This resistance mechanism is attributed to either a diminished uptake or an augmented efflux of MIL. Moreover, elevated expression of ABC transporters also contributes to resistance [93].

AmpB derived from *Streptomyces nodosus*, has been a stalwart in antileishmanial treatment, with a rich history spanning six to seven decades of use against fungal infections [94]. One of its distinguishing characteristics lies in its unique ability to selectively bind to ergosterol present in the cell membranes of *Leishmania* pathogen. This selective targeting sets it apart, as host cells predominantly contain cholesterol in their cell membranes, rendering the drug less sensitive to the host organism [95]. In the realm of leishmaniasis therapeutics, the specter of resistance looms large, even for stalwarts like AmpB. Instances of resistance have surfaced, particularly when the multidrug resistance MDR1 gene is found to be upregulated in resistant strains, indicating a heightened efflux of the drug [96]. Additionally, the involvement of ATP-binding cassette transporters, as well as amplification of the thiol pathway and alterations in membrane composition, have been implicated in promoting resistance within *Leishmania donovani* [96].

The evident vulnerability of monotherapy or single drug treatment to prompt resistance in leishmaniasis prompted researchers to introduce combination therapy [97]. This innovative approach involves administering limited doses of two drugs simultaneously. However, it was demonstrated that even combinations like MIL and paromomycin, as well as SSG and paromomycin, under experimental conditions, showed signs of resistance in both promastigotes and amastigotes [98].

# 1.7. Drug discovery approaches against Leishmaniasis

Various diseases pose significant challenges to public health, contributing to high mortality rates. In the realm of drug discovery, identifying effective drug targets for these diverse diseases becomes paramount in addressing the associated mortality. In the context of mortality associated with various diseases, drug discovery efforts aim to develop therapeutic interventions that can mitigate the impact of these conditions on human health [99]. Structural biology, system biology and bioinformatics can identify and validate potential drug targets which latter on leads to drug discovery [100]. Bioinformatics plays a crucial role in drug discovery by utilizing computational methods to analyze biological data, including genomics, proteomics, and other omics data [101]. Structural biology contributes to drug discovery by providing insights into the three-dimensional structures of biological molecules, such as proteins and enzymes [102]. Once identified, these drug targets become the focal point for designing and testing new drug molecules. The integration of bioinformatics, structural biology, and advanced computational approaches accelerates the drug discovery process, offering hope for more effective treatments and better outcomes for individuals affected by diverse diseases [103]. Drug targets are often proteins, that are involved in disease processes and can be modulated by drugs to achieve a therapeutic effect. Potential drug molecules are chemical compounds that have the potential to interact with these targets and exert a therapeutic effect.

# 1.8. Multitarget approach

The multitarget approach in drug development involves designing compounds or therapies that act on multiple targets or pathways within a biological system to achieve enhanced efficacy and therapeutic outcomes. This strategy acknowledges the complexity of diseases and aims to address various disease mechanisms simultaneously, thereby reducing the likelihood of resistance development and improving treatment effectiveness [104]. Observations have revealed that single-targeted drugs tend to develop resistance more rapidly compared to multitarget drugs. Over recent decades, an increasing number of pathogens have demonstrated resistance to specific antibiotics [105]. Antibiotics with dual or multiple targets exert their action through two mechanisms: firstly, by targeting closely related proteins within the same pathway, and secondly, by binding to distinct molecules involved in separate biological functions. This multifaceted approach enhances effectiveness and mitigates resistance development in pathogens [106]. Nevertheless, there are also numerous antibiotics that interact with two or more discrete molecules that are deemed essential for the treatment of certain diseases. Among the multitarget drugs utilized against diverse pathogens are penicillin, targeting penicillin-binding proteins [107]; ciprofloxacin, impacting both topoisomerase II and IV [108]; tetracycline, which acts on ribosomes and membranes [109]; polymyxin B, targeting both outer and inner membranes [110]; and tyrocidine, believed to affect the membrane and potentially DNA [111]. Miltefosine, a cornerstone in leishmaniasis treatment, is a prime example of a multitarget drug. It influences calcium homeostasis, the replication machinery, and mitochondrial functions, including cytochrome c oxidase activity and mitochondrial membrane integrity, while also impacting metacaspase production [112]. Cavalli and Bolognesi endeavored to identify fresh lead candidates for Leishmania and Trypanosoma utilizing multitarget ligands [113]. Overall, the multitarget approach shows considerable potential in treating a variety of diseases, including leishmaniasis, by potentially enhancing efficacy, minimizing the risk of resistance emergence, and ultimately improving patient outcomes.

# 1.9. Drug repurposing approach

Drug repurposing, represents an avenue of drug discovery wherein existing drugs, utilized for different therapeutic purposes, are reconsidered for novel indications, it is also considered as drug repositioning. It constitutes a retrospective approach, leveraging the established safety profiles and known mechanisms of action of these drugs to explore potential alternative targets for specific diseases [114]. Rather than creating entirely new medications from the ground up, drug repurposing capitalizes on the established safety profiles, pharmacokinetics, and other characteristics of existing drugs [115]. Repurposed drugs, having undergone preclinical and early-stage trials, decrease the risk of failure, particularly concerning safety in subsequent efficacy trials. This development process aids in reducing the time required for preclinical testing and ensures thorough safety assessments. [116]. Although investment requirements differ, there are potential cost savings in preclinical and early phase expenses. Although regulatory and later phase costs are more as

it was essential and crucial part similar to noble drug development, drug repurposing ultimately offers a more streamlined and cost-efficient method for introducing treatments for new indications [117].

Various computational and experimental methodologies, either independently or in synergy, contribute to the drug repurposing process. Computational strategies include signature matching, which compares specific drug characteristics with those of other drugs [118]; molecular docking, predicting binding interactions between a molecule and its target protein [119]; genome-wide association studies, identifying genes associated with a disease [120]; and pathway/network mapping, analyzing genetic and protein targets related to a disease [121]. Moreover, retrospective clinical analysis involves systematic scrutiny of electronic health records and clinical trial data [122], while novel data sources encompass large-scale *in vitro* drug screens coupled with genomic data and electronic health record databases [123]. Experimental approaches encompass phenotypic screening, employing *in vitro* or *in vivo* disease models [124], and binding assays utilizing techniques such as affinity chromatography and mass spectrometry to identify relevant target interactions.

Several successful examples of drug repurposing have been observed across various disease treatments. Zidovudine, initially employed in cancer therapy, was repurposed for HIV/AIDS treatment in 1987. Atomoxetine, originally indicated for Parkinson's disease, found utility in managing attention deficit hyperactivity disorder in 2002. Rituximab, utilized in cancer treatment, was later repurposed for Rheumatoid arthritis in 2006. Topiramate, initially prescribed for epilepsy, demonstrated efficacy in treating obesity in 2012. Ketoconazole, employed for fungal infections, was repurposed for Cushing syndrome in 2014. Sildenafil, initially intended for angina, was repurposed for erectile dysfunction in 1998 [117].

In the realm of drug discovery, Law et al. demonstrated the success of employing drug repurposing techniques, showcasing their effectiveness in picking out potential drug contender for treating a spectrum of diseases, ranging from cancers to influenza [125]. Recognizing the pressing challenge of drug resistance, Liu et al. emphasized a strategic solution—repurposing non-antibiotic drugs for diseases experiencing resistance. This approach not only holds promise for overcoming resistance but also lays the groundwork for the development of advanced combinational therapies, aiming to enhance efficacy and broaden treatment options [126]. Josef Jampilek pointed out that drug development is a complex process fraught with numerous obstacles, leading to a substantial failure rate and

prolonged timelines. In addressing this formidable challenge, repurposing existing drugs for antimicrobial purposes emerges as a promising strategy for effectively treating infections. This approach leverages established drugs to expedite the development of treatments for antimicrobial indications, potentially providing a more efficitive and practical different to traditional drug development methods [127]. Cheng et al. underscored the escalating challenge posed by multidrug-resistant (MDR) pathogens and emerging viruses. To effectively address this issue, they emphasized the importance of drug repurposing and drug combination screens. These strategies not only improve efficacy but also decrease the toxicity of combined drugs within the host [128]. Overall, drug repurposing represents a valuable strategy for maximizing the therapeutic potential of existing drugs, ultimately benefiting patients by providing innovative treatment options and potentially improving healthcare outcomes.

## 1.10. Subtractive genomics approach

Identifying drug targets represents a critical juncture in drug discovery. Leveraging the abundant genomic and proteomic data available across diverse sequence databases for both pathogens and hosts has streamlined the process of identifying drug targets for pathogens. Within this context, the subtractive genomic approach has emerged as a valuable tool. This bioinformatics methodology entails comparing genomes of distinct organisms, typically a pathogenic organism and its host or closely related counterparts, to discern genetic features unique to the pathogen [129]. At present, comparative and subtractive genomics are widely used to discover new targets. It aimed at crafting antimicrobial agents and vaccines against pathogens that are resistant to current therapies or lacking appropriate vaccine options. This approach aids in ascertaining the indispensability of these genes or proteins for pathogen survival and their absence in non-host homologues [130]. The overarching objective is to pinpoint potential drug targets or vaccine candidates specific to the pathogen, facilitating the development of targeted therapeutic strategies.

Recognizing the significance of this approach, George and Umrania reported its instrumental role in identifying possible therapeutic targets and uncovering drug-like compounds for *Streptococcus agalactiae*, *S. pneumoniae*, and *S. pyogenes*. This becomes particularly crucial in the face of emerging resistance observed in these bacterial strains [131]. Subtractive genomics has proven to be a dominant method for identifying new therapeutic drug targets, as evidenced by the comprehensive analysis of the entire proteome of

*Mycoplasma genitalium*. This process opens up avenues for designing potent drugs against the pathogen, highlighting the potential for innovative therapeutic interventions [132]. Recognizing the need to address Campylobacter infections in young children, Mehla and Ramana strategically employed subtractive genomics and metabolic pathway strategies. These approaches played a pivotal role in identifying novel drug targets against *Campylobacter jejuni*, the causative pathogen behind the infection, presenting a promising avenue for combating this health concern [133]. The subtractive genomic approach has been effectively utilized to identify new therapeutic targets and vaccine candidates for various pathogens, including bacteria, viruses, and parasites. It offers a systematic and efficient way to uncover unique features of pathogens that can be exploited for therapeutic intervention.

## **1.11.** Scope of the work

Despite decades of research, effective treatments for leishmaniasis are still limited, and the exposure of drug resistance has become a major challenge. The development of new drugs against L. donovani is imperative to combat the disease effectively and address the growing threat of drug resistance. With the ongoing emergence of resistance, the pathogenicity of Leishmaniasis is increasing, underscoring the need for new targets and potent ligands to counteract this trend. The scope of work on drug discovery for L. donovani and other Leishmania species underscores a multidisciplinary approach that integrates diverse scientific disciplines, encompassing multitarget strategies, drug repurposing, and subtractive genomics. The research endeavors to bridge gaps in our current understanding of drug development for leishmaniasis is to introduce innovative computational techniques. Our focus areas include identifying novel drug targets, screening compound libraries at high throughput, optimizing lead compounds, and assessing the binding affinity and stability of protein-ligand complexes. Our work focuses on five specific drug targets: adenine phosphoribosyl-transferase (APRT), a component of the purine pathway; dihydroorotate dehydrogenase (DHODH), involved in the pyrimidine pathway; pyridoxal kinase (PK), part of the vitamin B6 pathway; sterol alpha-14 demethylase (SDM), integral to sterol biosynthesis; and glutamate dehydrogenase (GDH). Throughout our study, we emphasize the meticulous selection of these protein targets and the application of computational techniques with diverse methodologies.

The central aim of the thesis is to advance and implement innovative methods that offer a more realistic and nuanced approach to target fishing and drug identification, particularly in

the context of a distinct class of diseases. By concentrating on leishmaniasis, the thesis contributes to the broader landscape of infectious diseases, offering insights and methodologies that may have implications for similar protozoan-driven ailments. The interdisciplinary nature of the research, combining various methods, underscores the commitment to advancing the field and potentially paving the way for more efficacious treatments for leishmaniasis. By enhancing our knowledge of parasite's biology and capitalizing on emerging therapeutic targets, we have the potential to create efficient treatments to alleviate the impact of leishmaniasis and enhance public health worldwide.

# Bibliography

- [1] Dallas, T. A., Laine, A. L. and Ovaskainen, O. Detecting parasite associations within multi-species host and parasite communities. *Proceedings of the Royal Society B*, 286(1912): 20191109, 2019.
- [2] García-Bernalt Diego, J., Fernandez-Soto, P. and Muro, A. LAMP in neglected tropical diseases: a focus on parasites. *Diagnostics*, 11(3):521, 2021.
- [3] Jones, C. M. and Welburn, S. C. Leishmaniasis beyond east Africa. *Frontiers in Veterinary Science*, 8:618766, 2021.
- [4] Rajkhowa, S., Hazarika, Z. and Jha, A. N. Systems biology and bioinformatics approaches in leishmaniasis. In *Applications of Nanobiotechnology for Neglected Tropical Diseases*, pages 509-548, Academic Press, 2021.
- [5] Kmetiuk, L. B., Tirado, T. C., Biondo, L. M., Biondo, A. W. and Figueiredo, F. B. Leishmania spp. in indigenous populations: A mini-review. *Frontiers in Public Health*, 10: 1033803, 2022.
- [6] Akhoundi, M., Kuhls, K., Cannet, A., Votýpka, J., Marty, P., Delaunay, P. and Sereno,
   D. A historical overview of the classification, evolution, and dispersion of
   Leishmania parasites and sandflies. *PLoS neglected tropical diseases*, 10(3):e0004349, 2016.
- [7] Serafim, T. D., Coutinho-Abreu, I. V., Dey, R., Kissinger, R., Valenzuela, J. G., Oliveira, F. and Kamhawi, S. Leishmaniasis: the act of transmission. *Trends in parasitology*, 37(11):976-987, 2021.
- [8] Kumar, R. and Nylén, S. Immunobiology of visceral leishmaniasis. *Frontiers in immunology*, 3:251, 2012.
- [9] Mundkur, S., Shashidhara, S., Hebbar, S. and Kanaparthi, S. Case 3: Hepatosplenomegaly with Hyperpigmentation in a 6-year-old Girl. *Pediatrics in Review*, 40(3):145-147, 2019.
- [10] Mans, D. R., Kent, A. D., Hu, R. V. P. F. and Schallig, H. D. F. H. Epidemiological, biological and clinical aspects of leishmaniasis with special emphasis on Busi Yasi in Suriname. *Journal of Clinical & Experimental Dermatology Research*, 8(2):e1000388, 2017.
- [11] Bezemer, J. M., Meesters, K., Naveda, C. L., Machado, P. R., Calvopiña, M., Leeflang, M. M., ... and de Vries, H. J. Clinical criteria for Mucosal leishmaniasis diagnosis in rural South America: a systematic literature review. *PLoS Neglected Tropical Diseases*, 16(8):e0010621, 2022.
- [12] Sabzevari, S., Mohebali, M. and Hashemi, S. A. Mucosal and mucocutaneous leishmaniasis in iran from 1968 to 2018: A narrative review of clinical features, treatments, and outcomes. *International Journal of Dermatology*, 59(5):606-612, 2020.
- [13] Ghatee, M. A., Taylor, W. R. and Karamian, M. The geographical distribution of cutaneous leishmaniasis causative agents in Iran and its neighboring countries, a review. *Frontiers in public health*, 8:11, 2020.

- [14] Salgado-Almario, J., Hernández, C. A. and Ovalle-Bracho, C. Geographical distribution of Leishmania species in Colombia, 1985-2017. *Biomédica*, 39(2):278-290, 2019.
- [15] WHO. Global Health Observatory Data Leishmaniasis. WHO. Available online at: http://www.who.int/gho/neglected\_diseases/leishmaniasis/en/ (accessed December 2023)
- [16] Barley, K., Mubayi, A., Safan, M. and Castillo-Chavez, C. A comparative assessment of visceral leishmaniasis burden in two eco-epidemiologically different countries, India and Sudan. *BioRxiv*, 592220:2019.
- [17] Patil, R. R. and Chatterjee, P. K. Epidemiology of Visceral Leishmaniasis in India, 2023.
- [18] Directorate National Vector borne disease control programme, Govt of India. Accelerated Plan for Kalazar Elimination. Directorate National Vector borne disease control programme, Govt of India; 2022.
- [19] Strazzulla, A., Cocuzza, S., Pinzone, M. R., Postorino, M. C., Cosentino, S., Serra, A., ... and Nunnari, G. Mucosal leishmaniasis: an underestimated presentation of a neglected disease. *BioMed Research International*, 2013:2013.
- [20] Pradhan, S., Schwartz, R. A., Patil, A., Grabbe, S. and Goldust, M. Treatment options for leishmaniasis. *Clinical and experimental dermatology*, 47(3):516-521, 2022.
- [21] Mathison, B. A. and Bradley, B. T. Review of the clinical presentation, pathology, diagnosis, and treatment of leishmaniasis. *Laboratory Medicine*, 54(4):363-371, 2023.
- [22] Gervazoni, L. F., Barcellos, G. B., Ferreira-Paes, T. and Almeida-Amaral, E. E. Use of natural products in leishmaniasis chemotherapy: an overview. *Frontiers in chemistry*, 1031, 2020.
- [23] Nico, D., Conde, L. and Palatnik de Sousa, C. B. Classical and modern drug treatments for leishmaniasis. *Antiprotozoal Drug Development and Delivery*, 1-21, 2021.
- [24] Haldar, A. K., Sen, P. and Roy, S. Use of antimony in the treatment of leishmaniasis: current status and future directions. *Molecular biology international*, 2011, 2011.
- [25] Aït-Oudhia, K., Gazanion, E., Vergnes, B., Oury, B. and Sereno, D. Leishmania antimony resistance: what we know what we can learn from the field. *Parasitology research*, 109:1225-1232, 2011.
- [26] Chávez-Fumagalli, M. A., Ribeiro, T. G., Castilho, R. O., Fernandes, S. O. A., Cardoso, V. N., Coelho, C. S. P., ... and Coelho, E. A. F. New delivery systems for amphotericin B applied to the improvement of leishmaniasis treatment. *Revista da Sociedade Brasileira de Medicina Tropical*, 48:235-242, 2015.
- [27] Handler, M. Z., Patel, P. A., Kapila, R., Al-Qubati, Y. and Schwartz, R. A. Cutaneous and mucocutaneous leishmaniasis: Differential diagnosis, diagnosis, histopathology, and management. *Journal of the American Academy of Dermatology*, 73(6):911-926, 2015.
- [28] Kumari, S., Kumar, V., Tiwari, R. K., Ravidas, V., Pandey, K. and Kumar, A. Amphotericin B: A drug of choice for Visceral Leishmaniasis. *Acta Tropica*, 106661, 2022.

- [29] Dorlo, T. P., Balasegaram, M., Beijnen, J. H. and de Vries, P. J. Miltefosine: a review of its pharmacology and therapeutic efficacy in the treatment of leishmaniasis. *Journal of Antimicrobial Chemotherapy*, 67(11):2576-2597, 2012.
- [30] Meheus, F., Balasegaram, M., Olliaro, P., Sundar, S., Rijal, S., Faiz, M. A. and Boelaert, M. Cost-effectiveness analysis of combination therapies for visceral leishmaniasis in the Indian subcontinent. *PLoS Neglected Tropical Diseases*, 4(9):e818, 2010.
- [31] Pokharel, P., Ghimire, R. and Lamichhane, P. Efficacy and safety of paromomycin for visceral leishmaniasis: A systematic review. *Journal of Tropical Medicine*, 2021, 2021.
- [32] Kaur, G. and Rajput, B. Comparative analysis of the omics technologies used to study antimonial, amphotericin B, and pentamidine resistance in leishmania. *Journal of parasitology research*, 2014, 2014.
- [33] Taslimi, Y., Zahedifard, F. and Rafati, S. Leishmaniasis and various immunotherapeutic approaches. *Parasitology*, 145(4):497-507, 2018.
- [34] Ikeogu, N. M., Akaluka, G. N., Edechi, C. A., Salako, E. S., Onyilagha, C., Barazandeh, A. F., & Uzonna, J. E. Leishmania immunity: advancing immunotherapy and vaccine development. *Microorganisms*, 8(8):1201, 2020.
- [35] Akbari, M., Oryan, A. and Hatam, G. Immunotherapy in treatment of leishmaniasis. *Immunology Letters*, 233:80-86, 2021.
- [36] Baxarias, M., Martínez-Orellana, P., Baneth, G., & Solano-Gallego, L. Immunotherapy in clinical canine leishmaniosis: a comparative update. *Research in veterinary science*, 125:218-226, 2019.
- [37] Ikeogu, N. M., Akaluka, G. N., Edechi, C. A., Salako, E. S., Onyilagha, C., Barazandeh, A. F. and Uzonna, J. E. Leishmania immunity: advancing immunotherapy and vaccine development. *Microorganisms*, 8(8):1201, 2020.
- [38] van Griensven, J. and Diro, E. Visceral leishmaniasis: recent advances in diagnostics and treatment regimens. *Infectious Disease Clinics*, 33(1):79-99, 2019.
- [39] Nafari, A., Cheraghipour, K., Sepahvand, M., Shahrokhi, G., Gabal, E. and Mahmoudvand, H. Nanoparticles: New agents toward treatment of leishmaniasis. *Parasite epidemiology and control*, 10:e00156, 2020.
- [40] Téllez, J., Echeverry, M. C., Romero, I., Guatibonza, A., Santos Ramos, G., Borges De Oliveira, A. C., ... and Demicheli, C. Use of liposomal nanoformulations in antileishmania therapy: Challenges and perspectives. *Journal of Liposome Research*, 31(2):169-176, 2021.
- [41] Vasile, C. Polymeric nanomaterials: Recent developments, properties and medical applications. *Polymeric nanomaterials in nanotherapeutics*, 1-66, 2019.
- [42] Gaspar, R., Préat, V., Opperdoes, F. R. and Roland, M. Macrophage activation by polymeric nanoparticles of polyalkylcyanoacrylates: activity against intracellular Leishmania donovani associated with hydrogen peroxide production. *Pharmaceutical research*, 9:782-787, 1992.
- [43] Abu Ammar, A., Nasereddin, A., Ereqat, S., Dan-Goor, M., Jaffe, C. L., Zussman, E. and Abdeen, Z. Amphotericin B-loaded nanoparticles for local treatment of cutaneous leishmaniasis. *Drug Delivery and Translational Research*, 9:76-84, 2019.

- [44] Shahnaz, G., Edagwa, B. J., McMillan, J., Akhtar, S., Raza, A., Qureshi, N. A., ... and Gendelman, H. E. Development of mannose-anchored thiolated amphotericin B nanocarriers for treatment of visceral leishmaniasis. *Nanomedicine*, 12(2):99-115, 2017.
- [45] Afzal, I., Sarwar, H. S., Sohail, M. F., Varikuti, S., Jahan, S., Akhtar, S. ... and Shahnaz, G. Mannosylated thiolated paromomycin-loaded PLGA nanoparticles for the oral therapy of visceral leishmaniasis. *Nanomedicine*, 14(4):387-406, 2019.
- [46] Ghasemiyeh, P., and Mohammadi-Samani, S. Solid lipid nanoparticles and nanostructured lipid carriers as novel drug delivery systems: Applications, advantages and disadvantages. *Research in pharmaceutical sciences*, 13(4):288-303, 2018.
- [47] Viegas, C., Patrício, A. B., Prata, J. M., Nadhman, A., Chintamaneni, P. K. and Fonte,
   P. Solid Lipid Nanoparticles vs. Nanostructured Lipid Carriers: A Comparative Review. *Pharmaceutics*, 15(6):1593, 2023.
- [48] Parvez, S., Yadagiri, G., Gedda, M. R., Singh, A., Singh, O. P., Verma, A., ... and Mudavath, S. L. Modified solid lipid nanoparticles encapsulated with Amphotericin B and Paromomycin: an effective oral combination against experimental murine visceral leishmaniasis. *Scientific Reports*, 10(1):12243, 2020.
- [49] Heidari-Kharaji, M., Taheri, T., Doroud, D., Habibzadeh, S., Badirzadeh, A. and Rafati, S. Enhanced paromomycin efficacy by solid lipid nanoparticle formulation against Leishmania in mice model. *Parasite immunology*, 38(10):599-608, 2016.
- [50] Allahverdiyev, A. M., Abamor, E. S., Bagirova, M. and Rafailovich, M. Antimicrobial effects of TiO2 and Ag2O nanoparticles against drug-resistant bacteria and leishmania parasites. *Future microbiology*, 6(8):933-940, 2011.
- [51] Abamor, E. S., Allahverdiyev, A. M., Bagirova, M. and Rafailovich, M. Meglumine antimoniate-TiO2@ Ag nanoparticle combinations reduce toxicity of the drug while enhancing its antileishmanial effect. *Acta tropica*, 169:30-42, 2017.
- [52] Allahverdiyev, A. M., Abamor, E. S., Bagirova, M., Ustundag, C. B., Kaya, C., Kaya, F. and Rafailovich, M. Antileishmanial effect of silver nanoparticles and their enhanced antiparasitic activity under ultraviolet light. International journal of Nanomedicine, 2705-2714, 2011.
- [53] Modabber, F. Vaccines against leishmaniasis. *Annals of Tropical Medicine & Parasitology*, 89(sup1):83-88, 1995.
- [54] Mayrink, W., Da Costa, C. A., Magalhães, P. A., Melo, M. N., Dias, M., Lima, A. O., ... and Williams, P. A field trial of a vaccine against American dermal leishmaniasis. *Transactions of the royal society of tropical medicine and hygiene*, 73(4): 385-387, 1979.
- [55] Machado-Pinto, J., Pinto, J., Da Costa, C. A., Genaro, O., Marques, M. J., Modabber, F. and Mayrink, W. Immunochemotherapy for cutaneous leishmaniasis: a controlled trial using killed Leishmania (Leishmania) amazonensis vaccine plus antimonial. *International journal of dermatology*, 41(2):73-78, 2002.
- [56] Armijos, R. X., Weigel, M. M., Romero, L., Garcia, V. and Salazar, J. Field trial of a vaccine against new world cutaneous leishmaniasis in an at-risk child population:

how long does protection last?. *The Journal of infectious diseases*, 187(12):1959-1961, 2003.

- [57] Convit, J., Ulrich, M., Zerpa, O., Borges, R., Aranzazu, N., Valera, M., ... and Tomedes, I. Immunotherapy of American cutaneous leishmaniasis in Venezuela during the period 1990–1999. *Transactions of the royal society of tropical medicine and hygiene*, 97(4):469-472, 2003.
- [58] Sharples, C. E., Shaw, M. A., Castes, M., Convit, J. and Blackwell, J. M. Immune response in healthy volunteers vaccinated with BCG plus killed leishmanial promastigotes: antibody responses to mycobacterial and leishmanial antigens. *Vaccine*, 12(15):1402-1412, 1994.
- [59] Momeni, A. Z., Jalayer, T., Emamjomeh, M., Khamesipour, A., Zicker, F., Ghassemi, R. L., ... and Modabber, F. A randomised, double-blind, controlled trial of a killed L. major vaccine plus BCG against zoonotic cutaneous leishmaniasis in Iran. *Vaccine*, 17(5):466-472, 1999.
- [60] Satti, I. N., Osman, H. Y., Daifalla, N. S., Younis, S. A., Khalil, E. A. G., Zijlstra, E. E., ... and Ghalib, H. W. Immunogenicity and safety of autoclaved Leishmania major plus BCG vaccine in healthy Sudanese volunteers. *Vaccine*, 19(15-16):2100-2106, 2001.
- [61] Coutinho De Oliveira, B., Duthie, M. S. and Alves Pereira, V. R. Vaccines for leishmaniasis and the implications of their development for American tegumentary leishmaniasis. *Human Vaccines & Immunotherapeutics*, 16(4):919-930, 2020.
- [62] Mitchell, G. F., Handman, E. and Spithill, T. W. Vaccination against cutaneous leishmaniasis in mice using nonpathogenic cloned promastigotes of Leishmania major and importance of route of injection. *Australian journal of experimental biology and medical science*, 62(2):145-153, 1984.
- [63] Gorczynski, R. M. Immunization of susceptible BALB/c mice against Leishmania braziliensis: II. Use of temperature-sensitive avirulent clones of parasite for vaccination purposes. *Cellular immunology*, 94(1):11-20, 1985.
- [64] Daneshvar, H., Coombs, G. H., Hagan, P. and Phillips, R. S. Leishmania mexicana and Leishmania major: attenuation of wild-type parasites and vaccination with the attenuated lines. *The journal of infectious diseases*, 187(10):1662-1668, 2003.
- [65] Dey, R., Dagur, P. K., Selvapandiyan, A., McCoy, J. P., Salotra, P., Duncan, R. and Nakhasi, H. L. Live attenuated Leishmania donovani p27 gene knockout parasites are nonpathogenic and elicit long-term protective immunity in BALB/c mice. *The Journal of Immunology*, 190(5):2138-2149, 2013.
- [66] Nagill, R. and Kaur, S. Vaccine candidates for leishmaniasis: a review. *International immunopharmacology*, 11(10):1464-1488, 2011.
- [67] Peacock, C. S., Seeger, K., Harris, D., Murphy, L., Ruiz, J. C., Quail, M. A., ... and Berriman, M. Comparative genomic analysis of three Leishmania species that cause diverse human disease. *Nature genetics*, 39(7):839-847, 2007.
- [68] Rachamim, N. and Jaffe, C. L. Pure protein from Leishmania donovani protects mice against both cutaneous and visceral leishmaniasis. *Journal of immunology* (*Baltimore, Md.: 1950*), 150(6):2322-2331, 1993.

- [69] Gillespie, P. M., Beaumier, C. M., Strych, U., Hayward, T., Hotez, P. J. and Bottazzi, M. E. Status of vaccine research and development of vaccines for leishmaniasis. *Vaccine*, 34(26):2992-2995, 2016.
- [70] Gomes, R., Oliveira, F., Teixeira, C., Meneses, C., Gilmore, D. C., Elnaiem, D. E., ... and Valenzuela, J. G. Immunity to sand fly salivary protein LJM11 modulates host response to vector-transmitted leishmania conferring ulcer-free protection. *Journal* of *Investigative Dermatology*, 132(12):2735-2743, 2012.
- [71] Duthie, M. S., Van Hoeven, N., Erasmus, J., Hsu, F. C. and Reed, S. G. Heterologous immunization with defined RNA and subunit vaccines enhances T cell responses that protect against Leishmania donovani. *Frontiers in immunology*, 9:410641, 2018.
- [72] Alarcon, J. B., Waine, G. W. and McManus, D. P. DNA vaccines: technology and application as anti-parasite and anti-microbial agents. *Advances in parasitology*, 42:343-410, 1999.
- [73] Rafati, S., Salmanian, A. H., Taheri, T., Vafa, M. and Fasel, N. A protective cocktail vaccine against murine cutaneous leishmaniasis with DNA encoding cysteine proteinases of Leishmania major. *Vaccine*, 19(25-26):3369-3375, 2001.
- [74] de Oliveira Gomes, D. C., Pinto, E. F., De Melo, L. D. B., Lima, W. P., Larraga, V., Lopes, U. G. and Rossi-Bergmann, B. Intranasal delivery of naked DNA encoding the LACK antigen leads to protective immunity against visceral leishmaniasis in mice. *Vaccine*, 25(12):2168-2172, 2007.
- [75] Melby, P. C., Yang, J., Zhao, W., Perez, L. E. and Cheng, J. Leishmania donovani p36 (LACK) DNA vaccine is highly immunogenic but not protective against experimental visceral leishmaniasis. *Infection and immunity*, 69(8):4719-4725, 2001.
- [76] Hobernik, D. and Bros, M. DNA vaccines—how far from clinical use?. *International journal of molecular sciences*, 19(11):3605, 2018.
- [77] Volpedo, G., Huston, R. H., Holcomb, E. A., Pacheco-Fernandez, T., Gannavaram, S., Bhattacharya, P., ... and Satoskar, A. R. From infection to vaccination: reviewing the global burden, history of vaccine development, and recurring challenges in global leishmaniasis protection. *Expert review of vaccines*, 20(11):1431-1446, 2021.
- [78] Vanaerschot, M., Dumetz, F., Roy, S., Ponte-Sucre, A., Arevalo, J. and Dujardin, J.
   C. Treatment failure in leishmaniasis: drug-resistance or another (epi-) phenotype?. *Expert review of anti-infective therapy*, 12(8):937-946, 2014.
- [79] Ponte-Sucre, A., Gamarro, F., Dujardin, J. C., Barrett, M. P., López-Vélez, R., García-Hernández, R., ... and Papadopoulou, B. Drug resistance and treatment failure in leishmaniasis: A 21st century challenge. *PLoS neglected tropical diseases*, 11(12): e0006052, 2017.
- [80] Ezra, N., Ochoa, M. T. and Craft, N. Human immunodeficiency virus and leishmaniasis. *Journal of global infectious diseases*, 2(3):248-257, 2010.
- [81] Cascio, A. and Colomba, C. Childhood Mediterranean visceral leishmaniasis. *Le Infezioni in Medicina*, 11(1):5-10, 2003.
- [82] Neal, R. A., Allen, S., McCoy, N., Olliaro, P. and Croft, S. L. The sensitivity of Leishmania species to aminosidine. *Journal of Antimicrobial Chemotherapy*, 35(5):577-584, 1995.

- [83] Fineman, M. S. and Augsburger, J. J. A new approach to an old problem. *Survey of ophthalmology*, 43(6):519-524, 1999.
- [84] Perry, M. R., Wyllie, S., Prajapati, V. K., Feldmann, J., Sundar, S., Boelaert, M. and Fairlamb, A. H. Visceral leishmaniasis and arsenic: an ancient poison contributing to antimonial treatment failure in the Indian subcontinent? *PLoS neglected tropical diseases*, 5(9):e1227, 2011.
- [85] Mandal, S., Maharjan, M., Singh, S., Chatterjee, M. and Madhubala, R. Assessing aquaglyceroporin gene status and expression profile in antimony-susceptible andresistant clinical isolates of Leishmania donovani from India. *Journal of Antimicrobial Chemotherapy*, 65(3):496-507, 2010.
- [86] Marquis, N., Gourbal, B., Rosen, B. P., Mukhopadhyay, R. and Ouellette, M. Modulation in aquaglyceroporin AQP1 gene transcript levels in drug-resistant Leishmania. *Molecular microbiology*, 57(6):1690-1699, 2005.
- [87] Manzano, J. I., García-Hernández, R., Castanys, S. and Gamarro, F. A new ABC halftransporter in Leishmania major is involved in resistance to antimony. *Antimicrobial* agents and chemotherapy, 57(8):3719-3730, 2013.
- [88] Perea, A., Manzano, J. I., Castanys, S., & Gamarro, F. The LABCG2 transporter from the protozoan parasite Leishmania is involved in antimony resistance. *Antimicrobial agents and chemotherapy*, 60(6):3489-3496, 2016.
- [89] Sundar, S., Makharia, A., More, D. K., Agrawal, G., Voss, A., Fischer, C., ... and Murray, H. W. Short-course of oral miltefosine for treatment of visceral leishmaniasis. *Clinical Infectious Diseases*, 31(4):1110-1113, 2000.
- [90] Paris, C., Loiseau, P. M., Bories, C. and Bréard, J. Miltefosine induces apoptosis-like death in Leishmania donovani promastigotes. *Antimicrobial agents and chemotherapy*, 48(3):852-859, 2004.
- [91] Sundar, S. and Olliaro, P. L. Miltefosine in the treatment of leishmaniasis: clinical evidence for informed clinical risk management. *Therapeutics and clinical risk management*, 3(5):733-740, 2007.
- [92] Pérez-Victoria, F. J., Sánchez-Cañete, M. P., Castanys, S. and Gamarro, F. Phospholipid translocation and miltefosine potency require both L. donovani miltefosine transporter and the new protein LdRos3 in Leishmania parasites. *Journal* of Biological Chemistry, 281(33): 23766-23775, 2006.
- [93] Castanys-Muñoz, E., Alder-Baerens, N., Pomorski, T., Gamarro, F. and Castanys, S. A novel ATP-binding cassette transporter from Leishmania is involved in transport of phosphatidylcholine analogues and resistance to alkyl-phospholipids. *Molecular microbiology*, 64(5):1141-1153, 2007.
- [94] Zhang, B., Zhou, Y. T., Jiang, S. X., Zhang, Y. H., Huang, K., Liu, Z. Q. and Zheng, Y. G. Amphotericin B biosynthesis in Streptomyces nodosus: quantitative analysis of metabolism via LC–MS/MS based metabolomics for rational design. *Microbial Cell Factories*, 19:1-12, 2020.
- [95] Lemke, A., Kiderlen, A. F. and Kayser, O. Amphotericin B. Appl Microbiol Biotechnol. 2005.

- [96] Purkait, B., Kumar, A., Nandi, N., Sardar, A. H., Das, S., Kumar, S. ... and Das, P. (2012). Mechanism of amphotericin B resistance in clinical isolates of Leishmania donovani. *Antimicrobial agents and chemotherapy*, 56(2):1031-1041.
- [97] van Griensven, J., Dorlo, T. P., Diro, E., Costa, C. and Burza, S. The status of combination therapy for visceral leishmaniasis: An updated review. *The Lancet Infectious Diseases*, 24(1):e36-e46, 2024.
- [98] Hendrickx, S., Beyers, J., Mondelaers, A., Eberhardt, E., Lachaud, L., Delputte, P., ...and Maes, L. Evidence of a drug-specific impact of experimentally selected paromomycin and miltefosine resistance on parasite fitness in Leishmania infantum. *Journal of Antimicrobial Chemotherapy*, 71(7):1914-1921, 2016.
- [99] Mohs, R. C., & Greig, N. H. Drug discovery and development: Role of basic biological research. Alzheimer's & Dementia: Translational Research & Clinical Interventions, 3(4): 651-657, 2017.
- [100] Sliwoski, G., Kothiwale, S., Meiler, J. and Lowe, E. W. Computational methods in drug discovery. *Pharmacological reviews*, 66(1):334-395, 2014.
- [101] Xia, X. Bioinformatics and drug discovery. *Current topics in medicinal chemistry*, 17(15):1709-1726, 2017.
- [102] Wishart, D. S. Bioinformatics in drug development and assessment. Drug metabolism reviews, 37(2):279-310, 2005.
- [103] Phoebe Chen, Y. P. and Chen, F. Identifying targets for drug discovery using bioinformatics. *Expert opinion on therapeutic targets*, 12(4):383-389, 2008.
- [104] Paolini, G. V., Shapland, R. H., van Hoorn, W. P., Mason, J. S. and Hopkins, A. L. Global mapping of pharmacological space. *Nature biotechnology*, 24(7):805-815, 2006.
- [105] Silver, L. L. Multi-targeting by monotherapeutic antibacterials. *Nature Reviews Drug Discovery*, 6(1):41-55, 2007.
- [106] Gray, D. A. and Wenzel, M. Multitarget approaches against multiresistant superbugs. *ACS infectious diseases*, 6(6):1346-1365, 2020.
- [107] Scheffers, D. J. and Pinho, M. G. Bacterial cell wall synthesis: new insights from localization studies. *Microbiology and molecular biology reviews*, 69(4):585-607, 2005.
- [108] Drlica, K., Malik, M., Kerns, R. J. and Zhao, X. Quinolone-mediated bacterial death. *Antimicrobial agents and chemotherapy*, 52(2):385-392, 2008.
- [109] Chopra, I. and Roberts, M. Tetracycline antibiotics: mode of action, applications, molecular biology, and epidemiology of bacterial resistance. *Microbiology and molecular biology reviews*, 65(2):232-260, 2001.
- [110] Teuber, M. and Bader, J. Action of polymyxin B on bacterial membranes: binding capacities for polymyxin B of inner and outer membranes isolated from Salmonella typhimurium G30. Archives of Microbiology, 109:51-58, 1976.
- [111] Bohg, A. and Ristow, H. Tyrocidine-induced modulation of the DNA conformation in Bacillus brevis. *European journal of biochemistry*, 170(1-2):253-258, 1987.
- [112] Braga, S. S. Multi-target drugs active against leishmaniasis: A paradigm of drug repurposing. *European Journal of Medicinal Chemistry*, 183:111660, 2019.

- [113] Cavalli, A. and Bolognesi, M. L. Neglected tropical diseases: multi-target-directed ligands in the search for novel lead candidates against Trypanosoma and Leishmania. *Journal of Medicinal Chemistry*, 52(23):7339-7359, 2009.
- [114] Medina-Franco, J. L., Giulianotti, M. A., Welmaker, G. S. and Houghten, R. A. Shifting from the single to the multitarget paradigm in drug discovery. *Drug discovery today*, 18(9-10):495-501, 2013.
- [115] Oprea, T. I. and Mestres, J. Drug repurposing: far beyond new targets for old drugs. *The AAPS journal*, 14:759-763, 2012.
- [116] Breckenridge, A. and Jacob, R. Overcoming the legal and regulatory barriers to drug repurposing. *Nature reviews Drug discovery*, 18(1):1-2, 2019.
- [117] Pushpakom, S., Iorio, F., Eyers, P. A., Escott, K. J., Hopper, S., Wells, A., ... and Pirmohamed, M. Drug repurposing: progress, challenges and recommendations. *Nature reviews Drug discovery*, 18(1):41-58, 2019.
- [118] Keiser, M. J., Setola, V., Irwin, J. J., Laggner, C., Abbas, A. I., Hufeisen, S. J., ... and Roth, B. L. Predicting new molecular targets for known drugs. *Nature*, 462(7270):175-181, 2009.
- [119] Saha, D., Borah, N. J. and Jha, A. N. Molecular scaffold recognition of drug molecules against essential genes of Leishmania donovani using biocomputing approach. *South African Journal of Botany*, 162:52-63, 2023.
- [120] Sanseau, P., Agarwal, P., Barnes, M. R., Pastinen, T., Richards, J. B., Cardon, L. R. and Mooser, V. Use of genome-wide association studies for drug repositioning. *Nature biotechnology*, 30(4):317-320, 2012.
- [121] Smith, S. B., Dampier, W., Tozeren, A., Brown, J. R. and Magid-Slav, M. Identification of common biological pathways and drug targets across multiple respiratory viruses based on human host gene expression analysis. *PloS* one, 7(3):e33174, 2012.
- [122] Ashburn, T. T. and Thor, K. B. Drug repositioning: identifying and developing new uses for existing drugs. *Nature reviews Drug discovery*, 3(8):673-683, 2004.
- [123] Huang, Y. H. and Vakoc, C. R. A biomarker harvest from one thousand cancer cell lines. *Cell*, 166(3):536-537, 2016.
- [124] Moffat, J. G., Vincent, F., Lee, J. A., Eder, J. and Prunotto, M. Opportunities and challenges in phenotypic drug discovery: an industry perspective. *Nature reviews Drug discovery*, 16(8):531-543, 2017.
- [125] Law, G. L., Tisoncik-Go, J., Korth, M. J. and Katze, M. G. Drug repurposing: a better approach for infectious disease drug discovery?. *Current opinion in immunology*, 25(5): 588-592, 2013.
- [126] Liu, Y., Tong, Z., Shi, J., Li, R., Upton, M. and Wang, Z. Drug repurposing for nextgeneration combination therapies against multidrug-resistant bacteria. *Theranostics*, 11(10): 4910, 2021.
- [127] Jampilek, J. Drug repurposing to overcome microbial resistance. *Drug Discovery Today*, 27(7):2028-2041, 2022.
- [128] Cheng, Y. S., Williamson, P. R. and Zheng, W. Improving therapy of severe infections through drug repurposing of synergistic combinations. *Current opinion in pharmacology*, 48:92-98, 2019.

- [129] Barh, D., Tiwari, S., Jain, N., Ali, A., Santos, A. R., Misra, A. N., ... and Kumar, A. In silico subtractive genomics for target identification in human bacterial pathogens. *Drug Development Research*, 72(2):162-177, 2011.
- [130] Akinnuwesi, A., Egieyeh, S. and Cloete, R. State-of-the-art strategies to prioritize Mycobacterium tuberculosis drug targets for drug discovery using a subtractive genomics approach. *Frontiers in Drug Discovery*, 3:1254656, 2023.
- [131] Georrge, J. J., & Umrania, V. V. (2012). Subtractive genomics approach to identify putative drug targets and identification of drug-like molecules for beta subunit of DNA polymerase III in Streptococcus species. *Applied biochemistry and biotechnology*, 167: 1377-1395.
- [132] Fatoba, A. J., Okpeku, M. and Adeleke, M. A. Subtractive genomics approach for identification of novel therapeutic drug targets in Mycoplasma genitalium. *Pathogens*, 10(8):921, 2021.
- [133] Mehla, K. and Ramana, J. Novel drug targets for food-borne pathogen Campylobacter jejuni: an integrated subtractive genomics and comparative metabolic pathway study. *Omics: a Journal of Integrative Biology*, 19(7):393-406, 2015.