## Dedicated to my beloved parents,

# Mr. Udoi Bhaskar Puzari (Deta) and

Mrs. Sangeeta Puzari (Maa)

for bestowing me with unconditional love, blessings, endless support and encouragement

In loving memory of my grandmother (Aita)

Late (Mrs.) Amiya Sharma

#### DECLARATION BY THE CANDIDATE

I hereby declare that the thesis "Development of Analytical Methods for Identification of Indian Snake Venoms and Indian Red Scorpion Venom" being submitted to Department of Molecular Biology and Biotechnology, Tezpur University, Tezpur, Assam in partial fulfillment for the award of the degree of Doctor of Philosophy in Molecular Biology and Biotechnology, has previously not formed the basis for the award of any degree, diploma, associateship, fellowship or any other similar title or recognition. Due to unavailability of proper facilities in Tezpur University, the following experiments/sample analyses were carried out at other institutes:

- LC-MS/MS analysis of protein samples were performed at National Centre for Cell Science, NCCS, Pune 411007, India.
- In vivo experiments and biophysical characterisation of gold nanoparticles are performed at Institute of Advanced Study in Science and Technology (IASST), Guwahati 781035, India

Date: 30/06/2025

Place: Tezpur

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#### **TEZPUR UNIVERSITY**

#### CERTIFICATE OF SUPERVISOR

This is to certify that the thesis entitled "Development of Analytical Methods for Identification of Indian Snake Venoms and Indian Red Scorpion Venom" submitted to the School of Sciences, Tezpur University in requirement of partial fulfilment for the award of the degree of Doctor of Philosophy in Molecular Biology and Biotechnology is a record of research work carried out by Ms. Upasana Puzari under my supervision and guidance. All help received by her from various sources have been duly acknowledged. No part of this thesis has been submitted elsewhere for award of any other degree.

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5.1	Determination of titer of purified antibody by ELISA: (a) (b) PAb 7, (c) PAb 8, and (d) PAb 9.	PAb 6, 135
	(e) Immune recognition of the individual purified PAbs tow respective CPs by dot blot assay; (f) Image analyses intensities of immune reactivity depicted by the PAbs. CPS and 4 denotes custom peptide 1, 2, 3 and 4. Error bars mean $\pm$ S.D (n=3).	of dot 3 1, 2, 3
5.2	(a) Immune-reactivity of the individual PAbs towards MTV 1:10, 1:20, 1:40, and 1:60 (MTV: PAb) determined by ELISA. Significance of difference of recognition by PAb 6 9 (1:4, 1:10, 1:20, 1:40, 1:60, MTV: PAb) compared to *p<0.05. Error bars indicate mean ± S.D. (n=3).	Indirect, 7, and
	(b) Comparison of the immune cross-reactivity of the four individually and in different combinations towards MTV (MTV: PAb) determined by Indirect ELISA. Significated difference of PAb individual and PAbF 2,3,4,5,6,7,8,9,10 compared to PAbF *p<0.05; MTV immune-recognition individual and PAbF 3,4,5,6,7,8 and 9 compared to PAbF 3,4,5,6,7,8 and 9 compared to PAbF 10, Ψp<0.05; MTV immune-recognition by PAb individual, 3,4,5,6,7,8 and 9 compared to PAbF 11, ωp<0.05. Error bars indicate mean ± S.D (n=3) bars indicate mean ± S.D (n=3). [PAbF 2 denotes PAb 6 (1:1:1:1, w/w/w/w), PAbF 3 denotes PAb 6+7 (1:1, w/w), denotes PAb 6+8 (1:1, w/w), PAbF 5 denotes PAb 6+9 (1:1, w/w), PAbF 6 denotes PAb 7+8 (1:1, w/w), PAbF 7 denotes PAb (1:1, w/w), PAbF 8 denotes PAb 8+9 (1:1, w/w), PAbF 9 denotes PAb 6+7+9 (1:1:1, w/w/w), PAbF 10 denotes PAb (1:1:1, w/w/w) and PAbF 11 denotes PAb 8+9+7 (1:1:1, w/w/w)	at 1:40 ance of and 11 by PAb AbF 2, PAbF mmune- pared to Error H7+8+9 PAbF 4 L, w/w), Ab 7+9
	(c) Comparison of the immune cross-reactivity between commercial ASA towards MTV determined by Indirect There is a significant difference in the fold change value the immune-reactivity of commercial ASA and PAbF at MTV doses, $*p < 0.05$ .	ELISA. petween

(d) Dot blot assay to determine immune-recognition of PAbs (individual PAb 1, 2, 3, 4, PAbF, PAbF 2, 3, 4, 5, 6, 7, 8, 9, 10, and 11) using anti-rabbit IgG-HRP and commercial ASA using anti-horse IgG-HRP; (e) Dot intensities of the immune-recognition demonstrated by the secondary antibodies as stated in (d). Error bars indicate mean  $\pm$  SD (n=3).

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(f) Dot blot assay of MTV (0.3 ng/ $\mu$ L) spiked rat plasma using the PAbs (individual PAb 1, 2, 3, and 4 and in combinations) and commercial ASA; (g) Image analyses of dot intensities of immune-reactivity determined as stated in (f). Significance of difference of PAb individual and PAbF 2,3,4,5,6,7,8,9, 10, 11 and commercial ASA compared to PAbF \*p<0.05; MTV immune-recognition by PAb individual and PAbF 2,3,4,5,6,7,8,9,10 and 11 compared to commercial ASA,  $^{\delta}$ p<0.05; MTV immune-recognition by PAb individual, PAbF 3,4,5,6,7,8,9,10 and 11 and commercial ASA compared to PAbF 2,  $^{\psi}$ p<0.05; MTV immune-recognition by PAb 7,8,9 and PAbF 3, 5, 7 compared to PAbF 10,  $^{\omega}$ p<0.05. Error bars indicate mean  $\pm$  S.D. (n=3).

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(h) Dot blot assay of MTV (0.3 ng/ $\mu$ L, 0.15 ng/ $\mu$ L and 0.075 ng/ $\mu$ L) spiked rat plasma using the PAbF. Blot 1 was incubated with MTV (0.075 ng/ $\mu$ L) spiked rat plasma; Blot 2 was incubated with MTV (0.15 ng/ $\mu$ L) spiked rat plasma, and Blot 3 was incubated with MTV (0.3 ng/ $\mu$ L) spiked rat plasma; (i) Image analyses of dot intensities were performed using ImageJ software. The dot intensities have been normalised against intensities of control without antigen. Significance of difference of 0.15 ng/ $\mu$ L dose of MTV compared to 0.3 ng/ $\mu$ L dose of MTV \*p<0.05. Error bars indicate mean  $\pm$  S.D. (n=3).

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(j) Western blot analysis to determine the immune recognition of MTV by PAbF. Immunoblot detected by HRP conjugated antirabbit IgG. (k) Western blot analysis to determine the immune recognition of MTV by Commercial ASA. Immunoblot detected by HRP conjugated anti-horse IgG. Lane MTV represents the immunoblot of MTV, and lane M denotes the marker. (l) Densitometry analyses of the blot intensities of MTV detected by PAbF and commercial ASA. Significance of difference in recognition of the MTV by PAbF compared to recognition by commercial ASA, \*p<0.05. Error bars indicate mean ± SD (n=3).

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(m)One-site specific binding curve representing spectrofluorometric interaction between a fixed concentration of MTV and graded concentrations of PAbF and commercial ASA (0.01 mg/mL, 0.02 mg/mL, 0.04 mg/mL, 0.08 mg/mL, 0.16 mg/mL, 0.32 mg/mL, 0.64 mg/mL, 1.28 mg/mL) showing the change in maximum fluorescence intensity ( $\lambda_{max}$ ) of MTV-PAbF and MTV-commercial ASA binding with a fixed concentration of MTV. The graphs were plotted using GraphPad Prism 5.0 software and shows the mean of five scans.

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5.3 (a) Dot blot assay comparing the immune-recognition of MTV in the plasma of the group I-IVrats by PAbF and commercial ASA when the blood was collected at 30 min, 60 min, and 120 min after the injection (i.v. and s.c.). Blots 1-3 incubated with control plasma (i.v.) collected after 30 min, 60 min, and 120 min recognised by PAbF; Blots 4-6 incubated with MTV-treated plasma (i.v.) collected after 30 min, 60 min, and 120 min recognised by PAbF; Blots 7-9 incubated with control plasma (s.c.) collected after 30 min, 60 min, and 120 min recognised by PAbF; Blots 10-12 incubated with MTV-treated plasma (s.c.) collected after 30 min, 60 min and 120 min recognised by PAbF; Blots 13-15 incubated with control plasma (s.c.) collected after 30 min, 60 min and 120 min recognised by commercial ASA; Blots 16-18 incubated with MTV-treated plasma (s.c.) collected after 30 min, 60 min and 120 min recognised by commercial ASA; (b) Image analyses of dot intensities of the group I-IV rats' plasma detection by PAbF and commercial ASA. The dot intensities have been normalised against intensities of control without antigen. Significance of difference in recognition of MTV-treated plasma collected at 30, 60, and 120 min by PAbF compared to recognition by commercial ASA \*p<0.05; recognition of MTV-treated plasma (s.c.) collected at 60 min and 120min PAbF compared to recognition of MTV-treated plasma collected at 30 min \(^{y}p<0.05\); recognition of MTV-treated plasma (s.c.) collected at 30 min and 120 min PAbF compared to recognition of MTV-treated plasma collected at 60 min <sup>\omega</sup>p<0.05; recognition of MTV-treated plasma (i.v.) collected at 30 min and 60 min by PAbF compared to recognition of MTV-treated plasma collected at 120 min  $^{\psi}p$ <0.05. Error bars indicate mean  $\pm$  S.D. (n=3).

(c) Comparison of immune cross-reactivity of PAbF towards MTV in the LMMPT-enriched MTV-treated and non-enriched plasma of envenomed rats. Blot 1 incubated with control non-enriched

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plasma; Blot 2 incubated with non-enriched MTV-treated plasma; Blot 3 incubated with control LMMPT-enriched MTV-treated plasma; Blot 4 incubated with LMMPT-enriched MTV-treated plasma; (d) Image analyses of dot intensities of the plasma detection by the PAbF. The dot intensities have been normalised against intensities of control without antigen. Significance of difference in recognition of LMMPT-enriched MTV-treated plasma compared to non-enriched MTV-treated plasma <sup>γ</sup>p<0.05; (e) Immune-reactivity of the PAbF towards MTV-treated non-enriched plasma and MTV-treated-LMMPT-enriched plasma determined by Sandwich ELISA. The absorbance values have been normalised against control without antigen. Significance of difference of recognition of MTV-treated-LMMPT-enriched plasma compared to MTV-treated non-enriched plasma \*p<0.05. Error bars indicate mean ± S.D. (n=3).

Multiple sequence alignments of the **(a)** K<sup>+</sup> channel toxin, and **(b)** Na<sup>+</sup> channel toxin identified by LC-MS/MS analysis and the MTV K<sup>+</sup> and Na<sup>+</sup> channel toxin used for designing the custom peptides.

(a) UV-Vis spectra of AuNP and AuNP conjugated with PAbF. The values are the mean of absorbance obtained in triplicates.
(b) FTIR spectrum of AuNP and AuNP conjugated with PAbF; (c)
Zeta potential of AuNP, AuNP functionalized with MUA, and

AuNP conjugated with PAbF.

5.5 TEM images of (d) AuNP and (e) AuNP- PAbF conjugate particle at 20 nm magnification; Particle size distribution histogram of (f) AuNP and (g) AuNP- PAbF conjugate particle in TEM images, with Gaussian function, fit using Originpro 8.5.

Topographic 2D AFM images with scanned area 1000 x 1000 nm of **(h)** AuNP, **(i)** AuNP-PAbF conjugate; Histogram of height distribution of **(j)** AuNP, **(k)** AuNP-PAbF conjugate, from the topographic 2D AFM images with scanned area 1000 x 1000 nm.

- (I) Calibration curve for estimating antibody (PAbF) left in the supernatant after conjugation to AuNP. Error bars indicate mean  $\pm$  S.D. (n=3).
- 5.6 **(a)** Absorbance spectra of the AuNP-PAbF conjugate in the presence of control (untreated rat plasma), MTV (0.3 ng/μL) spiked rat plasma, NnV, and RvV (50 ng/μL) spiked rat plasma. The

values are the mean of absorbance obtained in triplicates.

(b) Absorbance spectra of the AuNP-PAbF conjugate in the presence of control (untreated, group I and II) and MTV-treated-LMMPT-enriched plasma from group III and IV rats.

(c) Absorbance spectrum for MTV spiked rat plasma detection. Absorbance curves correspond to plasma samples containing 1-5 ng/μL MTV; (d) Calibration curve for MTV spiked rat plasma detection at concentrations 1-5 ng/μL. Error bars indicate mean ± S.D. (n=3).

## **ABBREVIATIONS**

Abbreviation	Full form
3D	Three-dimensional
3FTx	Three-finger toxin
20WBCT	20-min whole blood clotting test
AB-	Avidin-biotin micro enzyme-linked
microELISA	immunosorbent assay
AChE	Acetylcholinesterase
ACN	Acetonitrile
APase	Aminopeptidase
AFM	Atomic force microscopic
ANOVA	Analysis of variance
ASPro	Aspartic protease
ATP	Adenosine triphosphatase
ASA	Commercial equine anti-scorpion antivenom
AuNP	Gold nanoparticle
BLASTp	Protein-protein BLAST
BPP	Bradykinin potentiating peptide
BSA	Bovine serum albumin
β-BuTx	β-bungarotoxin
cDNA	Complementary Deoxyribonucleic Acid
ChE	Cholinesterase
CP	Custom peptide
CCSEA	Committee for Control and Supervision of Experiments on
	Animals
CMYK	Cyan, magenta, yellow and black
CRISP	Cysteine-rich secretory protein
CSL	Commonwealth Serum Laboratories
CTL	C-type lectin
CVF	Cobra venom factor
DMF	dimethyl formamide
DNase	Deoxyribonuclease
DTT	Dithiothreitol

Abbreviation	Full form
EcV	Echis carinatus venom
ECL	Enhanced Chemiluminescence
EDC	1-Ethyl-3-(3-dimethylaminopropyl) carbodiimide
EIA	Enzyme immunoassay
EIS	Electrochemical impedance spectroscopy
ELAA	Enzyme-linked aptamer assay
ELISA	Enzyme-linked immunosorbent assay
ExPASy	Expert Protein Analysis System
FELISA	Fluorogenic enzyme-linked immunosorbent assay
Fmoc	9-fluorenylmethoxycarbonyl
FPAb	Purified polyclonal snake venom toxin-specific antibody
	formulation
FTIR	Fourier-transform infrared spectroscopy
GC	Glutaminyl cyclase
GQDs	Graphene quantum dots
HAP	High-abundance protein
HCL	Hydrochloride
HMG CoA	Hydroxymethylglutaryl-coenzyme A
Нуа	Hyaluronidase
HRP	Horse radish peroxidise
HSS-Abs	Hemorrhagic species-specific antibodies
IgG	Immunoglobulin G
ISFET	Ion-sensitive field-effect transistor
KD	Dissociation constant
KLH	Keyhole Limpet Hemocyanin
KSPI	Kunitz-type proteinase inhibitor
KV	Bungarus caeruleus venom
LAAO	L-amino acid oxidase
LC-MS/MS	Liquid chromatography-tandem mass spectrometry
LFA	Lateral flow assay
LMMPT	Low molecular mass peptide toxins
LoD	Limit of detection

Abbreviation	Full form
LoQ	Limit of quantitation
LPP	Lipolysis potentiating peptides
LSPR	Localised surface plasmon resonance
mAb	Monoclonal antibody
MALDI-TOF	Matrix-assisted laser desorption/ionization -Time of flight
MBS	Maleimidobenzoyl-Nhydroxysuccinimide ester
MTV	Mesobuthus tamulus venom
MSI	Match Precursor Intensity
MUA	Mercaptoundecanoic acid
NCBI	National Center for Biotechnology Information
NEI	North-East India
NGF	Nerve growth factor
NHS	N-Hydroxysuccinimide
NnV	Naja naja venom
NkV	Naja kaouthia venom
NP	Natriuretic peptide
NSS-Abs	Neurotoxic species-specific antibodies
OIA	Optical immunoassay
OLP	Ohanin-like protein
PAb	Purified polyclonal antibody
PAbF	Purified polyclonal scorpion venom toxin-specific antibody
	formulation
PAV	Commercial equine anti-snake antivenom
PBS	Phosphate buffered saline
PCR	Polymerase chain reaction
PDB	Protein Data Bank
PDE	Phosphodiesterase
PIR	Protein Information Resource
PLA <sub>2</sub>	Phospholipase A <sub>2</sub>
PLB	Phospholipase B
PSVPL	Premium Serum and Vaccine Pvt. Ltd.
PVDF	Polyvinylidne fluoride

Abbreviation	Full form
Qdots	Quantum dots
RGB	Red, green and blue
<b>RGI-MDS</b>	Registrar General of India-Million Death Study
RIA	Radioimmunoassay
RP-HPLC	Reversed-phase high-performance liquid chromatography
RT-PCR	Reverse transcription polymerase chain reaction
RvV	Daboia russelii venom
SDS-PAGE	Sodium dodecyl sulfate-polyacrylamide gel electrophoresis
SELEX	Systematic Evolution of Ligands by EXponential
SPI	Serine protease inhibitor
SPLP	Serine protease-like protein
SPPS	Solid-phase peptide synthesis
SPR	Surface plasmon resonance
SSAbs	Species-specific antibodies
SVDK	Snake venom detection kit
SVMP	Snake venom metalloprotease
SVSP	Snake venom serine protease
SVTLE	Snake venom thrombin-like enzyme
TBS	Tris buffered saline
TBS-T	Tris buffered saline with 0.05% tween-20
TEM	Transmission electron microscope
TEMED	Tetramethylethylenediamine
$TMB/H_2O_2$	3,3,5,5'-tetramethylbenzidine/hydrogen peroxide
TiO <sub>2</sub>	Titanium dioxide
UniProtKB	Universal Protein Resource Knowledgebase
UV-Vis	Ultraviolet-visible
VDET	Venom Detection ELISA Test
VEGF	Vascular endothelial growth factor
Vesp	Vespryn
WHO	World health organization